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(54) Title: BIOSYNTHETIC BINDING PROTEIN	FOR C	CANCER MARKER
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(57) Abstract

Disclosed is a single-chain Fv (sFv) polypeptide defining a binding site which exhibits the immunological binding properties of an immunoglobulin molecule which binds c-erbB-2 or a c-erbB-2-related tumor antigen, the sFv includes at least two polypeptide domains connected by a polypeptide linker spanning the distance between the C-terminus of one domain and the N-terminus of the other, the amino acid sequence of each of the polypeptide domains includes a set of complementarity determining regions (CDRs) interposed between a set of framework regions (FRs), the CDRs conferring immunological binding to the c-erbB-2 or c-erbB-2-related tumor antigen.

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BIOSYNTHETIC BINDING PROTEIN FOR CANCER MARKER

This invention relates in general to novel biosynthetic compositions of matter and, specifically, to biosynthetic antibody binding site (BABS) proteins, and conjugates thereof. Compositions of the invention are useful, for example, in drug and toxin targeting, imaging, immunological treatment of various cancers, and in specific binding assays, affinity purification schemes, and biocatalysis.

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Background of the Invention

Carcinoma of the breast is the most common malignancy among women in North America, with 130,000 new cases in 1987. Approximately one in 11 women develop breast cancer in their lifetimes, causing this 15 malignancy to be the second leading cause of cancer death among women in the United States, after lung cancer. Although the majority of women with breast cancer present with completely resectable disease, 20 metastatic disease remains a formidable obstacle to The use of adjuvant chemotherapy or hormonal cure. therapy has definite positive impact on disease-free survival and overall survival in selected subsets of women with completely resected primary breast cancer, but a substantial proportion of women still relapse 25 with metastatic disease (see, e.g., Fisher et al. (1986) J. Clin. Oncol. 4:929-941; "The Scottish trial", Lancet (1987) 2:171-175). In spite of the regularly induced objective responses induced by chemotherapy and 30 hormonal therapy in appropriately selected patients, cure of metastatic breast cancer has not been achieved (see e.g., Aisner, et al. (187) J. Clin. Oncol.

<u>5</u>:1523-1533). To this end, many innovative treatment programs including the use of new agents, combinations of agents, high dose therapy (Henderson, <u>ibid.</u>) and increased dose intensity (Kernan et al. (1988) Clin.

- Invest. <u>259</u>:3154-3157) have been assembled. Although improvements have been observed, routine achievement of complete remissions of metastatic disease, the first step toward cure, has not occurred. There remains a pressing need for new approaches to treatment.
- The Fv fragment of an immunoglobulin molecule from IgM, and on rare occasions IgG or IgA, is produced by proteolytic cleavage and includes a non-covalent ${\rm V_{H}}^{-}$ ${\rm V_{L}}$ heterodimer representing an intact antigen binding site. A single chain Fv (sFv) polypeptide is a
- covalently linked $V_H^-V_L$ heterodimer which is expressed from a gene fusion including V_H^- and V_L^- encoding genes connected by a peptide-encoding linker. See Huston et al., 1988, Proc. Nat. Aca. Sci. 85: 5879, hereby incorporated by reference.
- U.S. Patent 4,753,894 discloses murine monoclonal antibodies which bind selectively to human breast cancer cells and, when conjugated to ricin A chain, exhibit a TCID 50% against at least one of MCF-7, CAMA-1, SKBR-3, or BT-20 cells of less than about 10 nM.
- The SKBR-3 cell line is recognized specifically by the monoclonal antibody 520C9. The antibody designated 520C9 is secreted by a murine hybridoma and is now known to recognize c-erbB-2 (Ring et al., 1991, Molecular Immunology 28:915).

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Summary of the Invention

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The invention features the synthesis of a class of novel proteins known as single chain Fv (sFv) polypeptides, which include biosynthetic single polypeptide chain binding sites (BABS) and define a binding site which exhibits the immunological binding properties of an immunoglobulin molecule which binds c-erbB-2 or a c-erbB-2-related tumor antigen.

The sFv includes at least two polypeptide domains connected by a polypeptide linker spanning the distance between the carboxy (C)- terminus of one domain and the amino (N)- terminus of the other domain, the amino acid sequence of each of the polypeptide domains including a set of complementarity determining regions (CDRs)

interposed between a set of framework regions (FRs), the CDRs conferring immunological binding to c-erbB-2 or a c-erbB-2 related tumor antigen.

In its broadest aspects, this invention features single-chain Fv polypeptides including biosynthetic antibody binding sites, replicable expression vectors prepared by recombinant DNA techniques which include and are capable of expressing DNA sequences encoding these polypeptides, methods for the production of these polypeptides, methods of imaging a tumor expressing c-erbB-2 or a c-erbB-2-related tumor antigen, and methods of treating a tumor using targetable therapeutic agents by virtue of conjugates or fusions with these polypeptides.

As used herein, the term "immunological binding"
or "immunologically reactive" refers to the noncovalent interactions of the type that occur between an
immunoglobulin molecule and an antigen for which the
immunoglobulin is specific; "c-erbB-2" refers to a

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protein antigen expressed on the surface of tumor cells, such as breast and ovarian tumor cells, which is an approximately 200,000 molecular weight acidic glycoprotein having an isoelectric point of about 5.3 5 and including the amino acid sequence set forth in SEQ ID NOS:1 and 2. A "c-erbB-2-related tumor antigen" is a protein located on the surface of tumor cells, such as breast and ovarian tumor cells, which is antigenically related to the c-erbB-2 antigen, i.e., bound by an immunoglobulin that is capable of binding the c-erbB-2 antigen, examples of such immunoglobulins being the 520C9, 741F8, and 454C11 antibodies; or which has an amino acid sequence that is at least 80% homologous, preferably 90% homologous, with the amino 15 acid sequence of c-erbB-2. An example of a c-erbB-2 related antigen is the receptor for epidermal growth factor.

An sFv CDR that is "substantially homologous with" an immunoglobulin CDR retains at least 70%, preferably 80% or 90%, of the amino acid sequence of the immunoglobulin CDR, and also retains the immunological binding properties of the immunoglobulin.

The term "domain" refers to that sequence of a polypeptide that folds into a single globular region in its native conformation, and may exhibit discrete binding or functional properties. The term "CDR" or complementarity determining region, as used herein, refers to amino acid sequences which together define the binding affinity and specificity of the natural Fv region of a native immunoglobulin binding site, or a synthetic polypeptide which mimics this function. CDRs typically are not wholly homologous to hypervariable regions of natural Fvs, but rather may also include specific amino acids or amino acid sequences which

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flank the hypervariable region and have heretofore been considered framework not directly determinative of complementarity. The term "FR" or framework region, as used herein, refers to amino acid sequences which are naturally found between CDRs in immunoglobulins.

Single-chain Fv polypeptides produced in accordance with the invention include biosyntheticallyproduced novel sequences of amino acids defining polypeptides designed to bind with a preselected 10 c-erbB-2 or related antigen material. The structure of these synthetic polypeptides is unlike that of naturally occurring antibodies, fragments thereof, or known synthetic polypeptides or "chimeric antibodies" in that the regions of the single-chain Fv responsible 15 for specificity and affinity of binding (analogous to native antibody variable (V_H/V_L) regions) may themselves be chimeric, e.g., include amino acid sequences derived from or homologous with portions of at least two different antibody molecules from the same or different species. These analogous V_H and V_{T} regions are connected from the N-terminus of one to the C-terminus of the other by a peptide bonded biosynthetic linker peptide.

The invention thus provides a single-chain Fv polypeptide defining at least one complete binding site capable of binding c-erbB-2 or a c-erbB-2-related tumor antigen. One complete binding site includes a single contiguous chain of amino acids having two polypeptide domains, e.g., V_H and V_L, connected by a amino acid linker region. An sFv that includes more than one complete binding site capable of binding a c-erbB-2-related antigen, e.g., two binding sites, will be a single contiguous chain of amino acids having four polypeptide domains, each of which is covalently linked

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by an amino acid linker region, e.g., $V_{\rm H1}$ -linker- $V_{\rm L1}$ -linker- $V_{\rm H2}$ -linker $V_{\rm L2}$. sFv's of the invention may include any number of complete binding sites ($V_{\rm H1}$ -linker- $V_{\rm L1}$), where n > 1, and thus may be a single contiguous chain of amino acids having n antigen binding sites and n X 2 polypeptide domains.

In one preferred embodiment of the invention, the single-chain Fv polypeptide includes CDRs that are substantially homologous with at least a portion of the amino acid sequence of CDRs from a variable region of an immunoglobulin molecule from a first species, and includes FRs that are substantially homologous with at least a portion of the amino acid sequence of FRs from a variable region of an immunoglobulin molecule from a second species. Preferably, the first species is mouse and the second species is human.

The amino acid sequence of each of the polypeptide domains includes a set of CDRs interposed between a set of FRs. As used herein, a "set of CDRs" refers to 3 CDRs in each domain, and a "set of FRS" refers to 4 FRs in each domain. Because of structural considerations, an entire set of CDRs from an immunoglobulin may be used, but substitutions of particular residues may be desirable to improve biological activity, e.g., based on observations of conserved residues within the CDRs of immunoglobulin species which bind c-erbB-2 related antigens.

In another preferred aspect of the invention, the CDRs of the polypeptide chain have an amino acid sequence substantially homologous with the CDRs of the variable region of any one of the 520C9, 741F8, and 454C11 monoclonal antibodies. The CDRs of the 520C9 antibody are set forth in the Sequence Listing as amino acid residue numbers 31 through 35, 50 through 66, 99

through 104, 159 through 169, 185 through 191, and 224 through 232 in SEQ ID NOS: 3 and 4, and amino acid residue numbers 31 through 35, 50 through 66, 99 through 104, 157 through 167, 183 through 189, and 222 through 230 in SEQ ID NOS: 5, and 6.

In one embodiment, the sFv is a humanized hybrid molecule which includes CDRs from the mouse 520C9 antibody interposed between FRs derived from one or more human immunoglobulin molecules. This hybrid sFv thus contains binding regions which are highly specific for the c-erbB-2 antigen or c-erbB-2-related antigens held in proper immunochemical binding conformation by human FR amino acid sequences, and thus will be less likely to be recognized as foreign by the human body.

In another embodiment, the polypeptide linker region includes the amino acid sequence set forth in the Sequence Listing as amino acid residue numbers 123 through 137 in SEQ ID NOS:3 and 4, and as amino acid residues 1-16 in SEQ ID NOS:11 and 12. In other embodiments, the linker sequence has the amino acid sequence set forth in the Sequence Listing as amino acid residues 121-135 in SEQ ID NOS:5 and 6, or the amino acid sequence of residues 1-15 in SEQ ID NOS:13 and 14.

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The single polypeptide chain described above also may include a remotely detectable moiety bound thereto to permit imaging or radioimmunotherapy of tumors bearing a c-erbB-2 or related tumor antigen. "Remotely detectable" moiety means that the moiety that is bound to the sFv may be detected by means external to and at a distance from the site of the moiety. Preferable remotely detectable moieties for imaging include radioactive atom such as "9" Technetium ("9" Tc), a gamma emitter. Preferable nucleotides for high dose

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radioimmunotherapy include radioactive atoms such as, (90 Yttrium (90 Yt), 131 Iodine (131 I) or 111 Indium (111 In).

In addition, the sFv may include a fusion protein

derived from a gene fusion, such that the expressed
sFv fusion protein includes an ancillary polypeptide
that is peptide bonded to the binding site polypeptide.
In some preferred aspects, the ancillary polypeptide
segment also has a binding affinity for a c-erbB-2 or
related antigen and may include a third and even a
fourth polypeptide domain, each comprising an amino
acid sequence defining CDRs interposed between FRs, and
which together form a second single polypeptide chain
biosynthetic binding site similar to the first
described above.

In other aspects, the ancillary polypeptide sequence forms a toxin linked to the N or C terminus of the sFv, e.g., at least a toxic portion of <u>Pseudomonas</u> exotoxin, phytolaccin, ricin, ricin A chain, or diphtheria toxin, or other related proteins known as ricin A chain-like ribosomal inhibiting proteins, i.e., proteins capable of inhibiting protein synthesis at the level of the ribosome, such as pokeweed antiviral protein, gelonin, and barley ribosomal protein inhibitor. In still another aspect, the sFv may include at least a second ancillary polypeptide or moiety which will promote internalization of the sFv.

The invention also includes a method for producing sFv, which includes the steps of providing a replicable expression vector which includes and which expresses a DNA sequence encoding the single polypeptide chain; transfecting the expression vector into a host cell to produce a transformant; and culturing the transformant to produce the sFv polypeptide.

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The invention also includes a m thod of imaging a tumor expressing a c-erbB-2 or related tumor antigen. This method includes the steps of providing an imaging agent including a single-chain Fv polypeptide as

5 described above, and a remotely detectable moiety linked thereto; administering the imaging agent to an organism harboring the tumor in an amount of the imaging agent with a physiologically-compatible carrier sufficient to permit extracorporeal detection of the

0 tumor; and detecting the location of the moiety in the subject after allowing the agent to bind to the tumor and unbound agent to have cleared sufficiently to permit visualization of the tumor image.

The invention also includes a method of treating cancer by inhibiting in vivo growth of a tumor expressing a c-erbB-2 or related antigen, the method including administering to a cancer patient a tumor inhibiting amount of a therapeutic agent which includes an sFv of the invention and at least a first moiety peptide bonded thereto, and which has the ability to limit the proliferation of a tumor cell.

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Preferably, the first moiety includes a toxin or a toxic fragment thereof, e.g., ricin A; or includes a radioisotope sufficiently radioactive to inhibit proliferation of the tumor cell, e.g., ⁹⁰Yt, ¹¹¹In, or ¹³¹I. The therapeutic agent may further include at least a second moiety that improves its effectiveness.

The clinical administration of the single-chain Fv or appropriate sFv fusion proteins of the invention, which display the activity of native, relatively small Fv of the corresponding immunoglobulin, affords a number of advantages over the use of larger fragments or entire antibody molecules. The single chain Fv and sFv fusion proteins of this invention offer fewer

cleavage sites to circulating proteolytic enzymes and thus offer greater stability. They reach their target tissue more rapidly, and are cleared more guickly from the body, which makes them ideal imaging agents for 5 tumor detection and ideal radioimmunotherapeutic agents for tumor killing. They also have reduced non-specific binding and immunogenicity relative to murine In addition, their expression from immunoglobulins. single genes facilitates targeting applications by 10 fusion to other toxin proteins or peptide sequences that allow specific coupling to other molecules or drugs. In addition, some sFv analogues or fusion proteins of the invention have the ability to promote the internalization of c-erbB-2 or related antigens 15 expressed on the surface of tumor cells when they are bound together at the cell surface. These methods permit the selective killing of cells expressing such antigens with the single-chain-Fv-toxin fusion of appropriate design. sFv-toxin fusion proteins of the invention possess 15-200-fold greater tumor cell 20 killing activity than conjugates which include a toxin that is chemically crosslinked to whole antibody or Fab.

Overexpression of c-erbB-2 or related receptors
on malignant cells thus allows targeting of sFv species
to the tumor cells, whether the tumor is well-localized
or metastatic. In the above cases, the internalization
of sFv-toxin fusion proteins permits specific
destruction of tumor cells bearing the over expressed
c-erbB-2 or related antigen. In other cases, depending
on the infected cells, the nature of the malignancy, or
other factors operating in a given individual, the same
c-erbB-2 or related receptors may be poorly
internalized or even represent a static tumor antigen

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population. In this event, the single-chain Fv and its fusion proteins can also be used productively, but in a different mode than applicable to internalization of the toxin fusion. Where c-erbB-2 receptor/sFv or sFv 5 fusion protein complexes are poorly internalized, toxins, such as ricin A chain, which operate cytoplasmically by inactivation of ribosomes, are not effective to kill cells. Nevertheless, single-chain unfused Fv is useful, e.g., for imaging or radioimmunotherapy, and bispecific single-chain Fv fusion proteins of various designs, i.e., that have two distinct binding sites on the same polypeptide chain, can be used to target via the two antigens for which the molecule is specific. For example, a bispecific single-chain antibody may have specificity for both the c-erbB-2 and CD3 antigens, the latter of which is present on cytotoxic lymphocytes (CTLs). bispecific molecule could thus mediate antibody dependent cellular cytotoxicity (ADCC) that results in

20 CTL-induced lysis of tumor cells. Similar results could be obtained using a bispecific single-chain Fv specific for c-erbB-2 and the Fcy receptor type I or Other bispecific sFv formulations include domains with c-erbB-2 specificity paired with a growth factor domain specific for hormone or growth factor receptors,

25 such as receptors for transferrin or epidermal growth factor (EGF).

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Brief Description of the Drawings

The foregoing and other objects of this invention, the various features thereof, as well as the invention itself, may be more fully understood from the following description, when read together with the accompanying drawings.

FIG. 1A is a schematic drawing of a DNA construct encoding an sFv of the invention, which shows the V_H and V_{τ} encoding domains and the linker region; FIG. 1B is a schematic drawing of the structure of Fv 10 illustrating $V_{_{\rm H}}$ and $V_{_{\rm T}}$ domains, each of which comprises three complementarity determining regions (CDRs) and four framework regions (FRs) for monoclonal 520C9, a well known and characterized murine monoclonal antibody specific for c-erbB-2; 15

FIGS. 2A-2E are schematic representations of embodiments of the invention, each of which comprises a biosynthetic single-chain Fv polypeptide which recognizes a c-erbB-2-related antigen: FIG. 2A is an sFv having a pendant leader sequence, FIG. 2B is an 20 sFv-toxin (or other ancillary protein) construct, and FIG. 2C is a bivalent or bispecific sFv construct; FIG. 2D is a bivalent sFv having a pendant protein attached to the carboxyl-terminal end; FIG. 2E is a bivalent sFv having pendant proteins attached to both amino- and carboxyl-terminal ends.

FIG. 3 is a diagrammatic representation of the construction of a plasmid encoding the 520C9 sFv-ricin A fused immunotoxin gene; and

FIG. 4 is a graphic representation of the results of a competition assay comparing the c-erbB-2 binding activity of the 520C9 monoclonal antibody (specific for c-erbB-2), an Fab fragment of that monoclonal antibody (filled dots), and different affinity purified

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fractions of the single-chain-Fv binding site for c-erbB-2 constructed from the variable regions of the 520C9 monoclonal antibody (sFv whole sample (+), sFv bound and eluted from a column of immobilized extracellular domain of C-erbB-2 (squares) and sFv flow-through (unbound, *)).

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Detailed Description of the Invention

Disclosed are single-chain Fv's and sFv fusion proteins having affinity for a c-erbB-2-related antigen expressed at high levels on breast and ovarian cancer 5 cells and on other tumor cells as well, in certain other forms of cancer. The polypeptides are characterized by one or more sequences of amino acids constituting a region which behaves as a biosynthetic antibody binding site. As shown in FIG. 1, the sites comprise heavy chain variable region (VH) 10, light chain variable region (V_{T_i}) 14 single chains wherein V_H 10 and V_T 14 are attached by polypeptide linker 12. The binding domains include CDRs 2, 4, 6 and 2', 4', 6' from immunoglobulin molecules able to bind a c-erbB-2related tumor antigen linked to FRs 32, 34, 36, 38 and 32', 34', 36' 38' which may be derived from a separate immunoglobulin. As shown in FIGS. 2A, 2B, and 2C, the BABS single polypeptide chains (V_H 10, V_T 14 and linker 12) may also include remotely detectable moieties and/or other polypeptide sequences 16, 18, or 22, which function e.g., as an enzyme, toxin, binding site, or site of attachment to an immobilization matrix or radioactive atom. Also disclosed are methods for producing the proteins and methods of their use.

The single-chain Fv polypeptides of the invention are biosynthetic in the sense that they are synthesized and recloned in a cellular host made to express a protein encoded by a plasmid which includes genetic sequence based in part on synthetic DNA, that is, a recombinant DNA made from ligation of plural, chemically synthesized and recloned oligonucleotides, or by ligation of fragments of DNA derived from the genome of a hybridoma, mature B cell clone, or a cDNA library derived from such natural sources.

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proteins of the invention are properly characterized as "antibody binding sites" in that these synthetic single polypeptide chains are able to refold into a 3-dimensional conformation designed specifically to 5 have affinity for a preselected c-erbB-2 or related tumor antigen. Single-chain Fv's may be produced as described in PCT application US88/01737, which corresponds to USSN 342,449, filed February 6, 1989, and claims priority from USSN 052,800, filed May 21, 1987, assigned to Creative BioMolecules, Inc., hereby incorporated by reference. The polypeptides of the invention are antibody-like in that their structure is patterned after regions of native antibodies known to be responsible for c-erbB-2-related antigen recognition.

More specifically, the structure of these biosynthetic antibody binding sites (BABS) in the region which imparts the binding properties to the protein, is analogous to the Fv region of a natural antibody to a c-erbB-2 or related antigen. It includes a series of regions consisting of amino acids defining at least three polypeptide segments which together form the tertiary molecular structure responsible for affinity and binding. The CDRs are held in appropriate conformation by polypeptide segments analogous to the framework regions of the Fv fragment of natural antibodies.

The CDR and FR polypeptide segments are designed empirically based on sequence analysis of the Fv region of preexisting antibodies, such as those described in U.S. Patent No. 4,753,894, herein incorporated by reference, or of the DNA encoding such antibody molecules.

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One such antibody, 520C9, is a murine monoclonal antibody that is known to react with an antigen expressed by the human breast cancer cell line SK-Br-3 (U.S. Patent 4,753,894). The antigen is an approximately 200 kD acidic glycoprotein that has an isoelectric point of 5.3, and is present at about 5 million copies per cell. The association constant measured using radiolabelled antibody is approximately 4.6 x 10⁸ M⁻¹.

10 In one embodiment, the amino acid sequences constituting the FRs of the single polypeptide chains are analogous to the FR sequences of a first preexisting antibody, for example, a human IgG. amino acid sequences constituting the CDRs are 15 analogous to the sequences from a second, different preexisting antibody, for example, the CDRs of a rodent or human IgG which recognizes c-erbB-2 or related antigens expressed on the surface of ovarian and breast tumor cells. Alternatively, the CDRs and FRs may be copied in their entirety from a single preexisting antibody from a cell line which may be unstable or, difficult to culture; e.g., an sFv-producing cell line that is based upon a murine, mouse/human, or human monoclonal antibody-secreting cell line.

Practice of the invention enables the design and biosynthesis of various reagents, all of which are characterized by a region having affinity for a preselected c-erbB-2 or related antigen. Other regions of the biosynthetic protein are designed with the particular planned utility of the protein in mind. Thus, if the reagent is designed for intravascular use in mammals, the FRs may include amino acid sequences that are similar or identical to at least a portion of the FR amino acids of antibodies native to that

mammalian species. On the other hand, the amino acid sequences that include the CDRs may be analogous to a portion of the amino acid sequences from the hypervariable region (and certain flanking amino acids) of an antibody having a known affinity and specificity for a c-erbB-2 or related antigen that is from, e.g., a mouse or rat, or a specific human antibody or immunoglobulin.

Other sections of native immunoglobulin protein structure, e.g., C_H and C_L, need not be present and normally are intentionally omitted from the biosynthetic proteins of this invention. However, the single polypeptide chains of the invention may include additional polypeptide regions defining a leader sequence or a second polypeptide chain that is bioactive, e.g., a cytokine, toxin, ligand, hormone, immunoglobulin domain(s), or enzyme, or a site onto which a toxin, drug, or a remotely detectable moiety, e.g., a radionuclide, can be attached.

One useful toxin is ricin, an enzyme from the 20 castor bean that is highly toxic, or the portion of ricin that confers toxicity. At concentrations as low as 1 ng/ml ricin efficiently inhibits the growth of cells in culture. The ricin A chain has a molecular 25 weight of about 30,000 and is glycosylated. ricin B chain has a larger size (about 34,000 molecular weight) and is also glycosylated. The B chain contains two galactose binding sites, one in each of the two domains in the folded subunit. The crystallographic structure for ricin shows the backbone tracing of the A 30 There is a cleft, which is probably the active site, that runs diagonally across the molecule. Also present is a mixture of «-helix, ß-structure, and irregular structure in the molecule.

The A chain enzymatically inactivates the 60S ribosomal subunit of eucaryotic ribosomes. The B chain binds to galactose-based carbohydrate residues on the surfaces of cells. It appears to be necessary to bind the toxin to the cell surface, and also facilitates and participates in the mechanics of entry of the toxin into the cell. Because all cells have galactose-containing cell surface receptors, ricin inhibits all types of mammalian cells with nearly the same efficiency.

Ricin A chain and ricin B chain are encoded by a gene that specifies both the A and B chains. polypeptide synthesized from the mRNA transcribed from the gene contains A chain sequences linked to B chain 15 sequences by a 'J' (for joining) peptide. peptide fragment is removed by post-translational modification to release the A and B chains. However, A and B chains are still held together by the interchain disulfide bond. The preferred form of ricin is 20 recombinant A chain as it is totally free of B chain and, when expressed in E. coli, is unglycosylated and thus cleared from the blood more slowly than the gycosylated form. The specific activity of the recombinant ricin A chain against ribosomes and that of 25 native A chain isolated from castor bean ricin are equivalent. An amino acid sequence and corresponding nucleic acid sequence of ricin A chain is set forth in the Sequence Listing as SEQ ID NOS:7 and 8.

Recombinant ricin A chain, plant-derived ricin A chain, deglycosylated ricin A chain, or derivatives thereof, can be targeted to a cell expressing a c-erbB-2 or related antigen by the single-chain Fv polypeptide of the present invention. To do this, the sFv may be chemically crosslinked to ricin A chain or

an active analog thereof, or in a preferred embodiment a single-chain Fv-ricin A chain immunotoxin may b formed by fusing the single-chain Fv polypeptide to one or more ricin A chains through the corresponding gene 5 fusion. By replacing the B chain of ricin with an antibody binding site to c-erbB-2 or related antigens, the A chain is guided to such antigens on the cell surface. In this way the selective killing of tumor cells expressing these antigens can be achieved. This selectivity has been demonstrated in many cases against cells grown in culture. It depends on the presence or absence of antigens on the surface of the cells to which the immunotoxin is directed.

The invention includes the use of humanized

single-chain-Fv binding sites as part of imaging
methods and tumor therapies. The proteins may be
administered by intravenous or intramuscular injection.
Effective dosages for the single-chain Fv constructs in
antitumor therapies or in effective tumor imaging can
be determined by routine experimentation, keeping in
mind the objective of the treatment.

The pharmaceutical forms suitable for injectable use include sterile aqueous solutions or dispersions. In all cases, the form must be sterile and must be fluid so as to be easily administered by syringe. It must be stable under the conditions of manufacture and storage, and must be preserved against the contaminating action of microorganisms. This may, for example, be achieved by filtration through a sterile 0.22 micron filter and/or lyophilization followed by sterilization with a gamma ray source.

Sterile injectable solutions are prepared by incorporating the single chain constructs of the invention in the required amount in the appropriate

solvent, such as sodium phosphate-buffered saline, followed by filter sterilization. As used herein, "a physiologically acceptable carrier" includes any and all solvents, dispersion media, antibacterial and antifungal agents that are non-toxic to humans, and the like. The use of such media and agents for pharmaceutically active substances is well known in the art. The media or agent must be compatible with maintenance of proper conformation of the single polypeptide chains, and its use in the therapeutic compositions. Supplementary active ingredients can also be incorporated into the compositions.

A bispecific single-chain Fv could also be fused to a toxin. For example, a bispecific sFv construct with specificity for c-erbB-2 and the transferrin receptor, a target that is rapidly internalized, would be an effective cytolytic agent due to internalization of the transferrin receptor/sFv-toxin complex. An sFv fusion protein may also include multiple protein domains on the same polypeptide chain, e.g., EGF-sFv-ricin A, where the EGF domain promotes internalization of toxin upon binding of sFv through interaction with the EGF receptor.

The single polypeptide chains of the invention

25 can be labelled with radioisotopes such as Iodine-131,

Indium-111, and Technetium-99m, for example. Beta
emitters such as Technetium-99m and Indium-111 are
preferred because they are detectable with a gamma
camera and have favorable half-lives for imaging in

30 vivo. The single polypeptide chains can be labelled,
for example, with radioactive atoms and as Yttrium-90,
Technetium-99m, or Indium-111 via a conjugated metal
chelator (see, e.g., Khaw et al. (1980) Science
209:295; Gansow et al., U.S. Patent No. 4,472,509;

Hnatowich, U.S. Patent No. 4,479,930), or by other standard means of isotope linkage to proteins known to those with skill in the art.

The invention thus provides intact binding sites for c-erbB-2 or related antigens that are analogous to $V_H^-V_L$ dimers linked by a polypeptide sequence to form a composite $(V_H^-linker^-V_L^-)_n$ or $(V_L^-linker^-V_H^-)_n$ polypeptide, where n is equal to or greater than 1, which is essentially free of the remainder of the antibody molecule, and which may include a detectable moiety or a third polypeptide sequence linked to each V_H^- or V_L^- .

FIGs. 2A-2E illustrate examples of protein structures embodying the invention that can be produced by following the teaching disclosed herein. All are characterized by at least one biosynthetic sFv single chain segment defining a binding site, and containing amino acid sequences including CDRs and FRs, often derived from different immunoglobulins, or sequences homologous to a portion of CDRs and FRs from different immunoglobulins.

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FIG. 2A depicts single polypeptide chain sFv 100 comprising polypeptide 10 having an amino acid sequence analogous to the heavy chain variable region (V_H) of a given anti-c-erbB-2 monoclonal antibody, bound through its carboxyl end to polypeptide linker 12, which in turn is bound to polypeptide 14 having an amino acid sequence analogous to the light chain variable region (V_L) of the anti-c-erbB-2 monoclonal. Of course, the light and heavy chain domains may be in reverse order. Linker 12 should be at least long enough (e.g., about 10 to 15 amino acids or about 40 Angstroms) to permit chains 10 and 14 to assume their proper conformation and interdomain relationship.

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Linker 12 may include an amino acid sequence homologous to a sequence identified as "self" by the species into which it will be introduced, if drug use is intended. Unstructured, hydrophilic amino acid sequences are preferred. Such linker sequences are set forth in the Sequence Listing as amino acid residue numbers 116 through 135 in SEQ ID NOS:3, 4, 5, and 6, which include part of the 16 amino acid linker sequences set forth in the Sequence Listing SEQ ID NOS:12 and 14.

Other proteins or polypeptides may be attached to either the amino or carboxyl terminus of protein of the type illustrated in FIG. 2A. As an example, leader sequence 16 is shown extending from the amino terminal end of $V_{\rm H}$ domain 10.

FIG. 2B depicts another type of reagent 200 including a single polypeptide chain 100 and a pendant protein 18. Attached to the carboxyl end of the polypeptide chain 100 (which includes the FR and CDR sequences constituting an immunoglobulin binding site) is a pendant protein 18 consisting of, for example, a toxin or toxic fragment thereof, binding protein, enzyme or active enzyme fragment, or site of attachment for an imaging agent (e.g., to chelate a radioactive ion such as Indium-111).

FIG. 2C illustrates single chain polypeptide 300 including second single chain polypeptide 110 of the invention having the same or different specificity and connected via peptide linker 22 to the first single polypeptide chain 100.

FIG. 2D illustrates single chain polypeptide 400 which includes single polypeptide chains 110 and 100 linked together by linker 22, and pendant protein 18 attached to the carboxyl end of chain 110.

FIG. 2E illustrates single polypeptide chain 500 which includes chain 400 of Fig. 2D and pendant protein 20 (EGF) attached to the amino terminus of chain 400.

As is evident from Figs. 2A-E, single chain

5 proteins of the invention may resemble beads on a

string by including multiple biosynthetic binding

sites, each binding site having unique specificity, or

repeated sites of the same specificity to increase the

avidity of the protein. As is evidenced from the

10 foregoing, the invention provides a large family of

reagents comprising proteins, at least a portion of

which defines a binding site patterned after the

variable region or regions of immunoglobulins to

c-erbB-2 or related antigens.

The single chain polypeptides of the invention are designed at the DNA level. The synthetic DNAs are then expressed in a suitable host system, and the expressed proteins are collected and renatured if necessary.

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The ability to design the single polypeptide chains of the invention depends on the ability to identify monoclonal antibodies of interest, and then to determine the sequence of the amino acids in the variable region of these antibodies, or the DNA sequence encoding them. Hybridoma technology enables production of cell lines secreting antibody to essentially any desired substance that elicits an immune response. For example, U.S. Patent No. 4,753,894 describes some monoclonal antibodies of interest which recognize c-erbB-2 related antigens on breast cancer cells, and explains how such antibodies were obtained. One monoclonal antibody that is particularly useful for this purpose is 520C9 (Bjorn et al. (1985) Cancer Res. 45:124-1221; U.S. Patent

No. 4,753,894). This antibody specifically recognizes the c-erbB-2 antigen expressed on the surface of various tumor cell lines, and exhibits very little binding to normal tissues. Alternative sources of sFv sequences with the desired specificity can take advantage of phage antibody and combinatorial library methodology. Such sequences would be based on cDNA from mice which were preimmunized with tumor cell membranes or c-erb-B-2 or c-erbB-2-related antigenic fragments or peptides. (See, e.g., Clackson et al, Nature 352 624-628 (1991))

The process of designing DNA that encodes the single polypeptide chain of interest can be accomplished as follows. RNA encoding the light and 15 heavy chains of the desired immunoglobulin can be obtained from the cytoplasm of the hyridoma producing the immunoglobulin. The mRNA can be used to prepare the cDNA for subsequent isolation of $\mathbf{V}_{\mathbf{H}}$ and $\mathbf{V}_{\mathbf{L}}$ genes by PCR methodology known in the art (Sambrook et al., 20 eds., Molecular Cloning, 1989, Cold Spring Harbor Laboratories Press, NY). The N-terminal amino acid sequence of H and L chain may be independently determined by automated Edman sequencing; if necessary, further stretches of the CDRs and flanking FRs can be 25 determined by amino acid sequencing of the H and L chain V region fragments. Such sequence analysis is now conducted routinely. This knowledge permits one to design synthetic primers for isolation of $\mathbf{V}_{\mathbf{H}}$ and $\mathbf{V}_{\mathbf{L}}$ genes from hybridoma cells that make monoclonal antibodies known to bind the c-erbB-2 or related antigen. These V genes will encode the Fv region that binds c-erbB-2 in the parent antibody.

Still another approach involves the design and construction of synthetic V genes that will encode an 5 Fv binding site specific for c-erbB-2 or related

receptors. For example, with the help of a computer program such as, for example, Compugene, and known variable region DNA sequences, one may design and directly synthesize native or near-native FR sequences from a first antibody molecule, and CDR sequences from a second antibody molecule. The V_H and V_L sequences described above are linked together directly via an amino acid chain or linker connecting the C-terminus of one chain with the N-terminus of the other.

These genes, once synthesized, may be cloned with or without additional DNA sequences coding for, e.g., a leader peptide which facilitates secretion or intracellular stability of a fusion polypeptide, or a leader or trailing sequence coding for a second polypeptide. The genes then can be expressed directly in an appropriate host cell.

By directly sequencing an antibody to a c-erbB-2 or related antigen, or obtaining the sequence from the literature, in view of this disclosure, one skilled in 20 the art can produce a single chain Fv comprising any desired CDR and FR. For example, using the DNA sequence for the 520C9 monoclonal antibody set forth in the Sequence Listing as SEQ ID NO:3, a single chain polypeptide can be produced having a binding affinity 25 for a c-erbB-2 related antigen. Expressed sequences may be tested for binding and empirically refined by exchanging selected amino acids in relatively conserved regions, based on observation of trends in amino acid sequence data and/or computer modeling techniques. 30 Significant flexibility in V_H and V_{T} design is possible because alterations in amino acid sequences may be made at the DNA level.

Accordingly, the construction of DNAs encoding the single-chain Fv and sFv fusion proteins of the

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invention can be done using known techniques involving the use of various restriction enzymes which make sequence-specific cuts in DNA to produce blunt ends or cohesive ends, DNA ligases, techniques enabling 5 enzymatic addition of sticky ends to blunt-ended DNA, construction of synthetic DNAs by assembly of short or medium length oligonucleotides, cDNA synthesis techniques, and synthetic probes for isolating immunoglobulin genes. Various promoter sequences and other regulatory RNA sequences used in achieving expression, and various type of host cells are also known and available. Conventional transfection techniques, and equally conventional techniques for cloning and subcloning DNA are useful in the practice of this invention and known to those skilled in the art. Various types of vectors may be used such as plasmids and viruses including animal viruses and bacteriophages. The vectors may exploit various marker genes which impart to a successfully transfected cell a detectable phenotypic property that can be used to identify which of a family of clones has successfully incorporated the recombinant DNA of the vector.

Of course, the processes for manipulating, amplifying, and recombining DNA which encode amino acid sequences of interest are generally well known in the art, and therefore, not described in detail herein. Methods of identifying the isolated V genes encoding antibody Fv regions of interest are well understood, and described in the patent and other literature. general, the methods involve selecting genetic material coding for amino acid sequences which define the CDRs and FRs of interest upon reverse transcription, according to the genetic code.

One method of obtaining DNA encoding the singlechain Fv disclosed herein is by assembly of synthetic oligonucleotides produced in a conventional, automated, polynucleotide synthesizer followed by ligation with 5 appropriate ligases. For example, overlapping, complementary DNA fragments comprising 15 bases may be synthesized semi-manually using phosphoramidite chemistry, with end segments left unphosphorylated to prevent polymerization during ligation. One end of the synthetic DNA is left with a "sticky end" corresponding 10 to the site of action of a particular restriction endonuclease, and the other end is left with an end corresponding to the site of action of another restriction endonuclease. Alternatively, this approach can be fully automated. The DNA encoding the single 15 chain polypeptides may be created by synthesizing longer single strand fragments (e.g., 50-100 nucleotides long) in, for example, a Biosearch oligonucleotide synthesizer, and then ligating the 20 fragments.

Additional nucleotide sequences encoding, for example, constant region amino acids or a bioactive molecule may also be linked to the gene sequences to produce a bifunctional protein.

25 For example, the synthetic genes and DNA fragments designed as described above may be produced by assembly of chemically synthesized oligonucleotides. 15-100mer oligonucleotides may be synthesized on a Biosearch DNA Model 8600 Synthesizer, and purified by 30 polyacrylamide gel electrophoresis (PAGE) in Tris-Borate-EDTA buffer (TBE). The DNA is then electroeluted from the gel. Overlapping oligomers may be phosphorylated by T4 polynucleotide kinase and ligated into larger blocks which may also be purified by PAGE.

The blocks or the pairs of longer oligonucleotides may be cloned in E. coli using a suitable cloning vector, e.g., pUC. Initially, this vector may be altered by single-strand mutagenesis to 5 eliminate residual six base altered sites. example, $V_{\rm H}$ may be synthesized and cloned into pUC as five primary blocks spanning the following restriction (1) EcoRI to first NarI site; (2) first NarI to XbaI; (3) XbaI to SalI; (4) SalI to NcoI; and (5) NcoI 10. to BamHI. These cloned fragments may then be isolated and assembled in several three-fragment ligations and cloning steps into the pUC8 plasmid. ligations, selected by PAGE, are then transformed into, for example, E. coli strain JM83, and plated onto LB 15 Ampicillin + Xgal plates according to standard procedures. The gene sequence may be confirmed by supercoil sequencing after cloning, or after subcloning into M13 via the dideoxy method of Sanger (Molecular Cloning, 1989, Sambrook et al., eds, 2d ed., Vol. 2, Cold Spring Harbor Laboratory Press, NY).

The engineered genes can be expressed in appropriate prokaryotic hosts such as various strains of $\underline{E.\ coli}$, and in eucaryotic hosts such as Chinese hamster ovary cells (CHO), mouse myeloma, hybridoma, transfectoma, and human myeloma cells.

If the gene is to be expressed in <u>E. coli</u>, it may first be cloned into an expression vector. This is accomplished by positioning the engineered gene downstream from a promoter sequence such as <u>Trp</u> or <u>Tac</u>, and a gene coding for a leader polypeptide such as fragment B (FB) of staphylococcal protein A. The resulting expressed fusion protein accumulates in refractile bodies in the cytoplasm of the cells, and may be harvested after disruption of the cells by

French press or sonication. The refractile bodies are solubilized, and the expressed fusion proteins are cleaved and refolded by the methods already established for many other recombinant proteins (Huston et al, 1988, supra) or, for direct expression methods, there is no leader and the inclusion bodies may be refolded without cleavage (Huston et al, 1991, Methods in Enzymology, vol 203, pp 46-88).

For example, subsequent proteolytic cleavage of
the isolated sFv from their leader sequence fusions can
be performed to yield free sFvs, which can be renatured
to obtain an intact biosynthetic, hybrid antibody
binding site. The cleavage site preferably is
immediately adjacent the sFv polypeptide and includes
one amino acid or a sequence of amino acids exclusive
of any one amino acid or amino acid sequence found in
the amino acid structure of the single polypeptide
chain.

The cleavage site preferably is designed for 20 specific cleavage by a selected agent. Endopeptidases are preferred, although non-enzymatic (chemical) cleavage agents may be used. Many useful cleavage agents, for instance, cyanogen bromide, dilute acid, trypsin, Staphylococcus aureus V-8 protease, postproline cleaving enzyme, blood coagulation Factor Xa, 25 enterokinase, and renin, recognize and preferentially or exclusively cleave at particular cleavage sites. One currently preferred peptide sequence cleavage agent is V-8 protease. The currently preferred cleavage site is at a Glu residue. Other useful enzymes recognize 30 multiple residues as a cleavage site, e.g., factor Xa (Ile-Glu-Gly-Arg) or enterokinase (Asp-Asp-Asp-Asp-Lys). Dilute acid preferentially leaves the peptide bond between Asp-Pro residues, and CNBr in acid cleaves after Met, unless it is followed by Tyr.

If the engineered gene is to be expressed in eucaryotic hybridoma cells, the conventional expression system for immunoglobulins, it is first inserted into an expression vector containing, for example, the immunoglobulin promoter, a secretion signal, immunoglobulin enhancers, and various introns. plasmid may also contain sequences encoding another polypeptide such as all or part of a constant region, enabling an entire part of a heavy or light chain to be 10 expressed, or at least part of a toxin, enzyme, cytokine, or hormone. The gene is transfected into myeloma cells via established electroporation or protoplast fusion methods. Cells so transfected may then express V_H -linker- V_L or V_L -linker- V_H single-chain Fv polypeptides, each of which may be attached in the 15 various ways discussed above to a protein domain having another function (e.g., cytotoxicity).

For construction of a single contiguous chain of amino acids specifying multiple binding sites, 20 restriction sites at the boundaries of DNA encoding a single binding site (i.e., V_H -linker- V_T) are utilized or created, if not already present. DNAs encoding single binding sites are ligated and cloned into shuttle plasmids, from which they may be further 25 assembled and cloned into the expression plasmid. order of domains will be varied and spacers between the domains provide flexibility needed for independent folding of the domains. The optimal architecture with respect to expression levels, refolding and functional activity will be determined empirically. To create 30 bivalent sFv's, for example, the stop codon in the gene encoding the first binding site is changed to an open reading frame, and several glycine plus serine codons including a restriction site such as BamHI (encoding

Gly-Ser) or XhoI (encoding Gly-Ser-Ser) are put in place. The second sFv gene is modified similarly at its 5' end, receiving the same restriction site in the same reading frame. The genes are combined at this site to produce the bivalent sFv gene.

Linkers connecting the C-terminus of one domain to the N-terminus of the next generally comprise hydrophilic amino acids which assume an unstructured configuration in physiological solutions and preferably are free of residues having large side groups which might interfere with proper folding of the V_H, V_L, or pendant chains. One useful linker has the amino acid sequence [(Gly)₄Ser]₃ (see SEQ ID NOS:5 and 6, residue numbers 121-135). One currently preferred linker has the amino acid sequence comprising 2 or 3 repeats of [(Ser)₄Gly], such as [(Ser)₄Gly]₂ and [(Ser)₄Gly]₃ (see SEQ ID NOS:3 and 4).

The invention is illustrated further by the following non-limiting Examples.

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EXAMPLES

1. Antibodies to c-erbB-2 Related Antigens

Monoclonal antibodies against breast cancer have been developed using human breast cancer cells or membrane extracts of the cells for immunizing mice, as described in Frankel et al. (1985) J. Biol. Resp. Modif. 4:273-286, hereby incorporated by reference. Hybridomas have been made and selected for production of antibodies using a panel of normal and breast cancer cells. A panel of eight normal tissue membranes, a fibroblast cell line, and frozen sections of breast cancer tissues were used in the screening. Candidates that passed the first screening were further tested on 16 normal tissue sections, 5 normal blood cell types,

11 nonbreast neoplasm sections, 21 breast cancer sections, and 14 breast cancer cell lines. From this selection, 127 antibodies were selected. Irrelevant antibodies and nonbreast cancer cell lines were used in control experiments.

Useful monoclonal antibodies were found to include 520C9, 454C11 (A.T.C.C. Nos. HB8696 and HB8484, respectively) and 741F8. Antibodies identified as selective for breast cancer in this screen reacted against five different antigens. The sizes of the 10 antigens that the antibodies recognize: 200 kD; a series of proteins that are probably degradation products with Mr's of 200 kD, 93kD, 60 kD, and 37 kD; 180 kD (transferrin receptor); 42 kD; and 55 kD, 15 respectively. Of the antibodies directed against the five classes of antigens, the most specific are the ones directed against the 200 kD antigen, 520C9 being a representative antibody for that antigen class. reacts with fewer breast cancer tissues (about 20-70% 20 depending on the assay conditions) and it reacts with the fewest normal tissues of any of the antibodies. 520C9 reacts with kidney tubules (as do many monoclonal antibodies), but not pancreas, esophagus, lung, colon, stomach, brain, tonsil, liver, heart, ovary, skin, 25 bone, uterus, bladder, or normal breast among some of the tissues tested.

2. <u>Preparation of cDNA Library Encoding 520C9</u> Antibody.

Polyadenylated RNA was isolated from

30 approximately 1 x 10⁸ (520C9 hybridoma) cells using the

"FAST TRACK" mRNA isolation kit from Invitrogen (San
Diego, CA). The presence of immunoglobulin heavy chain
RNA was confirmed by Northern analysis (Molecular
Cloning, 1989, Sambrook et al., eds., 2d ed., Cold

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Spring Harbor Laboratory Press, NY) using a recombinant probe containing the various J regions of heavy chain genomic DNA. Using 6 µg RNA for each, cDNA was prepared using the Invitrogen cDNA synthesis system with either random and oligo dT primers. Following synthesis, the cDNA was size-selected by isolating 0.5-3.0 Kilobase (Kb) fragments following agarose gel electrophoresis. After optimizing the cDNA to vector ratio, these fragments were then ligated to the pcDNA II Invitrogen cloning vector.

3. <u>Isolation of V_H and V_T Domains</u>

After transformation of the bacteria with plasmid library DNA, colony hybridization was performed using antibody constant (C) region and joining (J) region probes for either light or heavy chain genes. Orlandi, R., et al., 1989, Proc. Nat. Aca. Sci. 86:3833. The antibody constant region probe can be obtained from any of light or heavy chain nucleotide sequences from an immunoglobulin gene using known 20 procedures. Several potential positive clones were identified for both heavy and light chain genes and, after purification by a second round of screening, these were sequenced. One clone (M207) contained the sequence of non-functional Kappa chain which has a tyrosine substituted for a conserved cysteine, and also terminates prematurely due to a 4 base deletion which causes a frame-shift mutation in the variable-J region junction. A second light chain clone (M230) contained virtually the entire 520C9 light chain gene except for the last 18 amino acids of the constant region and approximately half of the signal sequence. The 520C9 heavy chain variable region was present on a clone of approximately 1,100 base pairs (F320) which ended near the end of the CH2 domain.

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4. Mutagenesis of V_H AND V_L

In order to construct the sFv, both the heavy and light chain variable regions were mutagenized to insert appropriate restriction sites (Kunkel, T.A., 1985, Proc. Nat. Acad. Sci. USA 82:1373). The heavy chain clone (F320) was mutagenized to insert a BamH1 site at the 5' end of V_H (F321). The light chain was also mutagenized simultaneously by inserting an EcoRV site at the 5' end and a PstI site with a translation stop codon at the 3' end of the variable region (M231).

5. Sequencing

cDNA clones encoding light and heavy chain were sequenced using external standard pUC primers and several specific internal primers which were prepared on the basis of the sequences obtained for the heavy chain. The nucleotide sequences were analyzed in a Genbank homology search (program Nucscan of DNA-star) to eliminate endogenous immunoglobulin genes. Translation into amino acids was checked with amino acid sequences in the NIH atlas edited by E. Kabat.

Amino acid sequences derived from 520C9 immunoglobulin confirmed the identity of these $V_{_{\mbox{\scriptsize H}}}$ and V_T cDNA clones. The heavy chain clone pF320 started 6 nucleotides upstream of the first ATG codon and 25 extended into the CH2-encoding region, but it lacked the last nine amino acid codons of the CH2 constant domain and all of the CH3 coding region, as well as the 3' untranslated region and the poly A tail. short heavy chain clone containing only the CH2 and CH3 coding regions, and the poly A tail was initially assumed to represent the missing part of the 52009 heavy chain. However, overlap between both sequences The 520C9 clone (pF320) encodes the was not identical. CH1 and CH2 domains of murine IgG1, whereas the short clone pF315 encodes the CH2 and CH3 of IgG2b.

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Gene Design

A nucleic acid sequence encoding a composite 520C9 sFv region containing a single-chain Fv binding site which recognizes c-erbB-2 related tumor antigens 5 was designed with the aid of Compugene software. gene contains nucleic acid sequences encoding the V, and V_{τ} regions of the 520C9 antibody described above linked together with a double-stranded synthetic oligonucleotide coding for a peptide with the amino acid sequence set forth in the Sequence Listing as amino acid residue numbers 116 through 133 in SEO ID NOS:3 and 4. This linker oligonucleotide contains helper cloning sites EcoRI and BamHI, and was designed to contain the assembly sites SacI and EcoRV near its 5' and 3' ends, respectively. These sites enable match-up and ligation to the 3' and 5' ends of 520C9 $V_{\rm H}$ and V_T , respectively, which also contain these sites $(V_H-linker-V_T)$. However, the order of linkage to the oligonucleotide may be reversed (V_L -linker- V_H) in this or any sFv of the invention. Other restriction sites were designed into the gene to provide alternative assembly sites. A sequence encoding the FB fragment of protein A was used as a leader.

The invention also embodies a humanized singlechain Fv, i.e., containing human framework sequences
and CDR sequences which specify c-erbB-2 binding, e.g.,
like the CDRs of the 520C9 antibody. The humanized Fv
is thus capable of binding c-erbB-2 while eliciting
little or no immune response when administered to a
patient. A nucleic acid sequence encoding a humanized
sFv may be designed and constructed as follows. Two
strategies for sFv design are especially useful. A
homology search in the GenBank database for the most
related human framework (FR) regions may be performed

and FR regions of the sFv may be mutagenized according to sequences identified in the search to reproduce the corresponding human sequence; or information from computer modeling based on x-ray structures of model Fab fragments may be used (Amit et al., 1986, Science 233:747-753; Colman et al., 1987, Nature 326:358-363; Sheriff et al., 1987, Proc. Nat. Aca. Sci., 84:8075-8079; and Satow et al., 1986, J. Mol. Biol. 190:593-604, all of which are hereby incorporated by In a preferred case, the most homologous reference). human $\boldsymbol{V}_{\boldsymbol{H}}$ and $\boldsymbol{V}_{\boldsymbol{L}}$ sequences may be selected from a collection of PCR-cloned human V regions. The FRs are made synthetically and fused to CDRs to make successively more complete V regions by PCR-based ligation, until the full humanized V_{τ} and V_{H} are 15 completed. For example, a humanized sFv that is a hybrid of the murine 520C9 antibody CDRs and the human myeloma protein NEW FRs can be designed such that each variable region has the murine binding site within a human framework (FR1-CDR1-FR2-CDR2-FR3-CDR3-FR4). Fab NEW crystal structure (Saul et al., 1978, J. Biol. Chem. 253:585-597) also may be used to predict the location of FRs in the variable regions. Once these regions are predicted, the amino acid sequence or the corresponding nucleotide sequence of the regions may be 25 determined, and the sequences may be synthesized and cloned into shuttle plasmids, from which they may be further assembled and cloned into an expression plasmid; alternatively, the FR sequences of the 520C9 sFv may be mutagenized directly and the changes verified by supercoil sequencing with internal primers

(Chen et al., 1985, DNA 4:165-170).

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7. <u>Preparation of and Purification 520C9 sFv</u>

A. Inclusion Body Solubilization.

The 520C9 sFv plasmid, based on a T₇ promoter and vector, was made by direct expression in <u>E. coli</u> of the fused gene sequence set forth in the Sequence Listing as SEQ. ID NO:3. Inclusion bodies (15.8 g) from a 2.0 liter fermentation were washed with 25 mM Tris, 10 mM EDTA, pH 8.0 (TE), plus 1 M guanidine hydrochloride (GuHCl). The inclusion bodies were solubilized in TE, 6 M GuHCl, 10 mM dithiothreitol (DTT), pH 9.0, and yielded 3825 A₂₈₀ units of material. This material was ethanol precipitated, washed with TE, 3M urea, then resuspended in TE, 8M urea, 10 mM DTT, pH 8.0. This precipitation step prepared the protein for ion exchange purification of the denatured sFv.

B. Ion Exchange Chromatography

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The solubilized inclusion bodies were subjected to ion exchange chromatography in an effort to remove contaminating nucleic acids and <u>E. coli</u> proteins before renaturation of the sFv. The solubilized inclusion bodies in 8M urea were diluted with TE to a final urea concentration of 6M, then passed through 100 ml of DEAE-Sepharose Fast Flow in a radial flow column. The sFv was recovered in the unbound fraction (69% of the starting sample).

The pH of this sFv solution (A₂₈₀ = 5.7; 290 ml) was adjusted to 5.5 with 1 M acetic acid to prepare it for application to an S-Sepharose Fast Flow column. When the pH went below 6.0, however, precipitate formed in the sample. The sample was clarified; 60% of the sample was in the pellet and 40% in the supernatant. The supernatant was passed through 100 ml S-Sepharose Fast Flow and the sFv recovered in the unbound fraction. The pellet was resolubilized in TE, 6 M

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GuHCl, 10 mm DTT, pH 9.0, and was also found to contain primarily sFv in a pool of 45 ml volume with an absorbance at 280 nm of 20 absorbance units. This reduced sFv pool was carried through the remaining steps of the purification.

C. Renaturation of sFv

Renaturation of the sFv was accomplished using a disulfide-restricted refolding approach, in which the disulfides were oxidized while the sFv was fully denatured, followed by removal of the denaturant and refolding. Oxidation of the sFv samples was carried out in TE, 6 M GuHCl, 1 mM oxidized glutathione (GSSG), 0.1 mM reduced glutathione (GSH), pH 9.0. The sFv was diluted into the oxidation buffer to a final protein A₂₈₀ = 0.075 with a volume of 4000 ml and incubated overnight at room temperature. After overnight oxidation this solution was dialyzed against 10 mM sodium phosphate, 1 mM EDTA, 150 mM NaCl, 500 mM urea, pH 8.0 (PENU) [4 x (20 liters X 24 hrs)]. Low levels of activity were detected in the refolded sample.

Membrane Fractionation and Concentration of Active sFv

In order to remove aggregated misfolded material before any concentration step, the dialyzed refolded 520C9 sFv (5050 ml) was filtered through a 100K MWCO membrane (100,000 mol. wt. cut-off) (4 x 60 cm²) using a Minitan ultrafiltration device (Millipore). This step required a considerable length of time (9 hours), primarily due to formation of precipitate in the retentate and membrane fouling as the protein concentration in the retentate increased. 95% of the protein in the refolded sample was retained by the 100K membranes, with 79% in the form of insoluble material. The 100K retentate had very low activity and was discarded.

The 100K filtrate contained most of the soluble sFv activity for binding c-erbB-2, and it was next concentrated using 10K MWCO membranes (10,000 mol. wt. cut-off) (4 x 60 cm²) in the Minitan, to a volume of 100 ml (50X). This material was further concentrated using a YM10 10K MWCO membrane in a 50 ml Amicon stirred cell to a final volume of 5.2 ml (1000X). Only a slight amount of precipitate formed during the two 10K concentration steps. The specific activity of this concentrated material was significantly increased relative to the initial dialyzed refolding.

E. Size Exclusion Chromatography of Concentrated sFv

When refolded sFv was fractionated by size
exclusion chromatography, all 520C9 sFv activity was
determined to elut at the position of folded monomer.
In order to enrich for active monomers, the 1000X
concentrated sFv sample was fractionated on a Sephacryl
S-200 HR column (2.5 x 40 cm) in PBSA (2.7 mM KCl, 1.1

mm KH₂PO₄, 138 mm NaCl, 8.1 mm Na₂HPO₄ 7H₂O, 0.02%
NaN₃) + 0.5 M urea. The elution profile of the column
and SDS-PAGE analysis of the fractions showed two sFv
monomer peaks. The two sFv monomer peak fractions were
pooled (10 ml total) and displayed c-erbB-2 binding
activity in competition assays.

F. Affinity Purification of 520C9 sFv

The extracellular domain of (ECD) c-erbB-2 was
expressed in bacculovirus-infected insect cells. This
protein (ECD c-erbB-2) was immobilized on an agarose
30 affinity matrix. The sFv monomer peak was dialyzed
against PBSA to remove the urea and then applied to a
0.7 x 4.5 cm ECD c-erbB-2-agarose affinity column in
PBSA. The column was washed to baseline A₂₈₀, then
eluted with PBSA + 3 M LiCl, pH = 6.1. The peak

fractions were pooled (4 ml) and dialyzed against PBSA to remove the LiCl. 72 μ g of purified sFv was obtained from 750 μq of S-200 monomer fractions. Activity measurements on the column fractions were determined by 5 a competitive assay. Briefly, sFv affinity purification fractions and HRP-conjugated 520C9 Fab fragments were allowed to compete for binding to SK-BR-3 membranes. Successful binding of the sFv preparation prevented the HRP-52069 Fab fragment from 10 binding to the membranes, thus also reducing or preventing utilization of the HRP substrate, and no color development (see below for details of competition assay). The results showed that virtually all of the sFv activity was bound by the column and was recovered in the eluted peak (Figure 4). As expected, the specific activity of the eluted peak was increased relative to the column sample, and appeared to be essentially the same as the parent Fab control, within the experimental error of these measurements.

20 9. Yield After Purification.

Table I shows the yield of various 520C9 preparations during the purification process. Protein concentration (μg/ml) was determined by the BioRad protein assay. Under "Total Yield", 300 AU denatured sFv stock represents 3.15 g inclusion bodies from 0.4 liters fermentation. The oxidation buffer was 25 mM Tris, 10 mM EDTA, 6 M GdnHCl, 1 MM GSSG, 0.1 mM GSH, pH 9.0. Oxidation was performed at room temperature overnight. Oxidized sample was dialyzed against 10 mM sodium phosphate, 1 mM EDTA, 150 mM NaCl, 500 mM urea, pH 8.0. All subsequent steps were carried out in this buffer, except for affinity chromatography, which was carried out in PBSA.

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Table I

5	<u>Sample</u>	<u>Volume</u>	Protein Concentration	Total Yield X	Yield
3	1. Refolding III (oxidation)	4000 ml	0.075 A ₂₈₀	300 AU	-
10	2. Dialyzed Refolding III	5050 ml	38 μg/ml	191.9 mg	100
15	3. Minitan 100K Filtrate	5000 ml	2 μg/ml	10.0 mg	5.4
13	4. Minitan 10K Retentate	100 ml	45 μg/ml	4.5 mg	2.3
20	6. YM10 10K Retentate	5.2 ml	600 μg/ml	3.1 mg	1.6
	7. S-200 sFv Monomer Peak	10.0 ml	58 μg/ml	0.58 mg	0.3
25	8. Affinity Purified sFv	5.5 ml	13 μg/ml	0.07 mg	0.04

10. Immunotoxin Construction

The ricin A-520C9 single chain fused immunotoxin (SEQ. ID NO:7) encoding gene was constructed by isolating the gene coding for ricin A on a HindIII to 5 BamH1 fragment from pPL229 (Cetus Corporation, Emeryville, CA) and using it upstream of the 520C9 sFv in pH777, as shown in FIG. 3. This fusion contains the 122 amino acid natural linker present between the A and B domains of ricin. However, in the original pRAP229 expression vector the codon for amino acid 268 of ricin was converted to a TAA translation stop codon so that the expression of the resulting gene produces only ricin A. Therefore, in order to remove the translation stop codon, site-directed mutagenesis was performed to remove the TAA and restore the natural serine codon. This then allows translation to continue through the entire immunotoxin gene.

In order to insert the immunotoxin back into the pPL229 and pRAP229 expression vectors, the PstI site at the end of the immunotoxin gene had to be converted to a sequence that was compatible with the BamHI site in vector. A synthetic oligonucleotide adaptor containing a BclI site nested between PstI ends was inserted. BclI and BamHI ends are compatible and can be combined into a hybrid BclI/BamHI site. Since BclI nuclease is sensitive to dam methylation, the construction first was transformed into a dam(-) E. coli strain, Gm48, in order to digest the plasmid DNA with BclI (and HindIII), then insert the entire immunotoxin gene on a HindIII/BclI fragment back into both Hind III/BamHI-digested expression vectors.

When native 520C9 IgG1 is conjugated with native ricin A chain or recombinant ricin A chain, the resulting immunotoxin is able to inhibit protein

synthesis by 50% at a conc ntration of about 0.4 x 10^{-9} M against SK-Br-3 cells. In addition to reacting with SK-Br-3 breast cancer cells, native 520C9 IgG1 immunotoxin also inhibits an ovarian cancer cell line, OVCAR-3, with a ID₅₀ of 2.0 x 10^{-9} M.

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In the ricin A-sFv fusion protein described above, ricin acts as leader for expression, i.e., is fused to the amino terminus of sFv. Following direct expression, soluble protein was shown to react with antibodies against native 520C9 Fab and also to exhibit ricin A chain enzymatic activity.

In another design, the ricin A chain is fused to the carboxy terminus of sFv. The 520C9 sFv may be secreted via the PelB signal sequence with ricin A chain attached to the C-terminus of sFv. For this construct, sequences encoding the PelB-signal sequence, sFv, and ricin are joined in a bluescript plasmid via a HindIII site directly following sFv (in our expression plasmids) and the HindIII site preceding the ricin gene, in a three part assembly (RI-HindIII-BamHI). new PstI site following the ricin gene is obtained via the Bluescript polylinker. Mutagenesis of this DNA removes the stop codon and the original PstI site at the end of sFv, and places several serine residues between the sFv and ricin genes. This new gene fusion, PelB signal sequence/sFv/ricin A, can be inserted into expression vectors as an EcoRI/PstI fragment.

In another design, the pseudomonas exotoxin fragment analogous to ricin A chain, PE40, is fused to the carboxy terminus of the anti-c-erbB-2 741F8 sFv (Seq ID NOS: 15 and 16). The resulting 741F8 sFv-PE40 is a single-chain Fv-toxin fusion protein, which was constructed with an 18 residue short FB leader which initially was left on the protein. E. coli expression

of this protein produced inclusion bodies that were refolded in a 3 M urea glutathione/redox buffer. resulting sFv-PE40 was shown to specifically kill c-erbB-2 bearing cells in culture more fully and with 5 apparently better cytotoxicity than the corresponding crosslinked immunotoxin. The sFv-toxin protein, as well as the 741F8 sFv, can be made in good yields by these procedures, and may be used as therapeutic and diagnostic agents for tumors bearing the c-erbB-2 or 10 related antigens, such as breast and ovarian cancer.

11. Assays

A. Competition ELISA

SK-Br-3 extract is prepared as a source of c-erbB-2 antigen as follows. SK-Br-3 breast cancer cells (Ring et al. 1989, Cancer Research 49:3070-3080), are grown to near confluence in Iscove's medium (Gibco BRL, Gaithersburg, Md.) plus 5% fetal bovine serum and 2 mM glutamine. The medium is aspirated, and the cells are rinsed with 10 ml fetal bovine serum (FBS) plus 20 calcium and magnesium. The cells are scraped off with a rubber policeman into 10 ml FBS plus calcium and magnesium, and the flask is rinsed out with another 5 ml of this buffer. The cells are then centrifuged at 100 rpm. The supernate is aspirated off, and the cells are resuspended at 10⁷ cells/ml in 10 mM NaCl, 0.5% NP40, pH 8 (TNN buffer), and are pipetted up and down to dissolve the pellet. The solution is then centrifuged at 1000 rpm to remove nuclei and other insoluble debris. The extract is filtered through 0.45 Millex HA and 0.2 Millex Gv filters. The TNN extract is stored as aliquots in Wheaton freezing vials at -70°C.

A fresh vial of SK-Br-3 TNN extract is thawed and diluted 200-fold into deionized water. Immediately thereafter, 40ug per well are added to a Dynatech PVC 35

96 well plak, which is allowed to sit overnight in a 37°C dry incubator. The plates are washed four times in phosphate buffered saline (PBS), 1% skim milk, 0.05% Tween 20.

The non-specific binding sites are blocked as follows. When the plate is dry, 100 ug per well PBS is added containing 1% skim milk, and the incubation allowed to proceed for one hour at room temperature.

The single-chain Fv test samples and standard

520C9 whole antibody dilutions are then added as
follows. 520C9 antibody and test samples are diluted
in dilution buffer (PBS + 1% skim milk) in serial twofold steps, initially at 50ug/ml and making at least 10
dilutions for 520C9 standards. A control containing
only dilution buffer is included. The diluted samples
and standards are added at 50ul per well and incubated
for 30 minutes at room temperature.

The 520C9-horseradish peroxidase (HRP) probe is added as follows. 520C9-HRP conjugate (Zymed Labs., South San Francisco, California) is diluted to 14 ug/ml 20 with 1% skim milk in dilution buffer. The optimum dilutions must be determined for each new batch of peroxidase conjugate without removing the previous 20 ul per well of probe was added and incubated The plate is then 25 for one hour at room temperature. washed four times in PBS. The peroxidase substrate is The substrate solution should be made then added. fresh for each use by diluting tetramethyl benzidine stock (TMB; 2mg/ml in 100% ethanol) 1:20 and 3% hydrogen peroxide stock 1:2200 in substrate buffer 30 (10mM sodium acetate, 10mM Na, EDTA, pH 5.0). incubated for 30 minutes at room temperature. wells are then quenched with 100 ul per well 0.8 M H₂SO₄ and the absorbance at 150 nm read.

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refolded but unpurified 520C9 monoclonal antibody,
520C9 Fab fragments, and the 520C9 sFv single-chain
binding site after binding and elution from an affinity
column (eluted) or the unbound flow through fraction
(passed). In Fig. 4, the fully purified 520C9 sFv
exhibits an affinity for c-erbB-2 that is
indistinguishable from the parent monoclonal antibody,
within the error of measuring protein concentration.

B. In vivo testing

Immunotoxins that are strong inhibitors of protein synthesis against breast cancer cells grown in culture may be tested for their in vivo efficacy. The in vivo assay is typically done in a nude mouse model using xenografts of human MX-1 breast cancer cells. Mice are injected with either PBS (control) or different concentrations of sFv-toxin immunotoxin, and a concentration-dependent inhibition of tumor growth will be observed. It is expected that higher doses of immunotoxin will produce a better effect.

The invention may be embodied in other specific forms without departing from the spirit and scope thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather than by the foregoing description, and all changes which come within the meaning and range of equivalence of the claims are intended to be embraced therein.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: Huston, James S.
 Oppermann, Hermann
 Houston, L. L.
 Ring, David B.
- (ii) TITLE OF INVENTION: Biosynthetic Binding Protein for Cancer Marker
- (iii) NUMBER OF SEQUENCES: 16
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 - (C) CITY: Boston
 - (D) STATE: Massachusetts
 - (E) COUNTRY: USA
 - (F) ZIP: 02109
- (V) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: Pitcher, Edmund R.
 - (B) REGISTRATION NUMBER: 27,829
 - (C) REFERENCE/DOCKET NUMBER: 2054/22
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- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 4299 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A).
- (B)
- NAME/KEY: CDS LOCATION: 1..4299 OTHER INFORMATION: /note= "product = "c-erb-b-2"" (D)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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CCC Pro	CCC	GGA Gly	GCC Ala 20	GCG Ala	AGC Ser	ACC Thr	CAA Gln	GTG Val 25	TGC Cys	ACC Thr	GGC Gly	ACA Thr	GAC Asp 30	ATG Met	AAG Lys	•	96
CTG Leu	CGG Arg	CTC Leu 35	CCT Pro	GCC Ala	AGT Ser	CCC Pro	GAG Glu 40	ACC Thr	CAC His	CTG Leu	GAC Asp	ATG Met 45	CTC Leu	CGC Arg	CAC His		144
CTC Leu	TAC Tyr 50	CAG Gln	GGC Gly	TGC Cys	CAG Gln	GTG Val 55	GTG Val	CAG Gln	GGA Gly	AAC Asn	CTG Leu 60	GAA Glu	CTC Leu	ACC Thr	TAC Tyr	•	192
CTG Leu 65	CCC Pro	ACC Thr	AAT Asn	GCC Ala	AGC Ser 70	CTG Leu	TCC Ser	TTC Phe	CTG Leu	CAG Gln 75	GAT Asp	ATC Ile	CAG Gln	GAG Glu	GTG Val 80		240
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CAG Gln	AGG Arg	CTG Leu	CGG Arg 100	ATT Ile	GTG Val	CGA Arg	GGC Gly	ACC Thr 105	CAG Gln	CTC Leu	TTT Phe	GAG Glu	GAC Asp 110	AAC Asn	TAT Tyr		336
GCC Ala	CTG Leu	GCC Ala 115	Val	CTA	GAC Asp	AAT Asn	GGA Gly 120	GAC Asp	CCG Pro	CTG Leu	AAC Asn	AAT Asn 125	ACC Thr	ACC Thr	CCT Pro		384
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CTC Leu 145	Thr	GAG Glu	ATC	TTG Leu	AAA Lys 150	Gly	GGG Gly	GTC Val	TTG Leu	ATC Ile 155	GIn	CGG Arg	AAC Asn	CCC	CAG Gln 160		480
CTC Leu	TGC Cys	TAC	CAG Gln	GAC Asp 165	Thr	ATT	TTG Leu	TGG	AAG Lys 170	Asp	ATC	TTC Phe	CAC His	AAG Lys 175	AAC Asn		528
AAC Asn	CAG Gln	CTG Leu	GCT Ala 180	Leu	ACA Thr	CTG Leu	ATA	GAC Asp 185	Thr	AAC Asn	CGC	TCT Ser	CGG Arg	Ala	TGC Cys		576

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	TGT Cys							672
	AAG Lys							720
	TGC Cys							768
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	ACA Thr							864
	GGC Gly							912
	GTG Val							960
	GCA Ala							1008
	CGA Arg 340							1056
	GTT Val							1104
	GGG Gly							1152
	AAC Asn							1200
	GAA Glu							1248

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TTC TGC Cys 595 TCC TAC ATG CCC ATC Pro Cys Pro 11e	Asp Ser Leu Pro Asp 420 GGA CGA ATT CTG CAC Gly Arg 11e Leu His 435 GGC ATC AGC TGG CTG Trp Leu 450 CTG GCC CTC ATC CAC His 465 CCC TGG GAC CAG CTC Leu Ala Leu 11e His 485 GCC AAC CGG CCA GAG ARG Ala Asn Arg 515 GTC AAC CTG TGC GCC CGA Arg Ser Gln 530 CGA GTA CTG CAG GGG Arg Val Leu Gln Gly 545 TTG CCG TGC TGC CAC CCT Leu Pro Cys His Pro 565 TTT GGA CCG TGC CAC CCT Phe Gly Pro Glu Ala 580 CCT CCC TTC TGC GTG CTC CTC CTC CTC TGC CTC CT	Asp Ser Leu Pro Asp Leu 420 GGA CGA ATT CTG CAC AAT Leu His Asn 435 GGC ATC AGC TGG CTG GGG Trp Leu Gly 450 CTG GCC CTC ATC CAC CAT His 470 CCC TGG GAC CAC CAC CAT His 470 CCC TGG GAC CAC CAC CAT His 485 GCC AAC CGG CCA GAG GAC AAT ASp 500 CAG CTG TGC GCC CGA GGG GIN Leu Cys Ala Arg Gly 515 GTC AAC TGC Cys ATC CAG CTC TTC Phe 530 CGA GTA CTG CAG CAG CAG TTC Cys Ser Gln Phe 530 TTG CCG TGC CAC CAC CAC CAC ATC ASp 580 CCT CCC TTC TGC GAG GCC CAC CAC CAC CAC CAC CAC CAC C	ASP SET LEU PRO ASP LEU SET 420 GGA CGA ATT CLEU HIS ASN GLY GGC ATC AGC TGG CTG GGG CTG GLY ILE SET Trp LEU GLY LEU 455 CTG GCC CTC ATC CAC CAT AAC LEU ALA LEU ILE HIS Phe Arg 485 GCC TGG GAC CAG CTC TTT CGG PRO TTP ASP CLEU Phe ARG 485 GCC AAC CGG CCA GAG GAC GAC ALA ASP CLU ASP	Asp Ser Leu Pro Asp Leu Ser Val GGA CGA ATT CTG CAC AAT GGC GCC Gly Arg lile Leu His Asn Gly Ala 440 GGC ATC AGC TGG CTG GGG CTG CGC Gly Ile Ser Trp Leu Gly Leu Arg 450 CTG GCC TTC ATC Leu His His Asn ACC Leu Ala Leu Ile His His Asn Thr 465 CCC TGG GAC CAG CTG TTT CGG AAC Pro Trp Asp Gln Leu Phe Arg Asn GCC AAC AGG CCA GAG GAC GAC TGC AAC Asn Arg Pro Glu Asp Glu Cys 500 CAG CTG TGC GCC CGA GGG CAC TGC Gln Leu Cys Ala Arg Gly His Cys 515 CTC AAC TGC AGC CAG GGG CTC CTG AASn Cys Ser Gln Phe Leu Arg 530 CGA GTA CTG CAG GGG CTC CCT AGG Arg Val Leu Gln Gly Leu Pro Arg 545 TTG CCG TGC CAC CAC GAG GGG CTC CCC AGG Arg Cys Fro Glu Ala Asp Gln Cys TTT GAA CCG GAG GCT GAC CAG Leu Pro Cys His Pro Glu Cys Cln TTT GAA CCG GAG GCT CCS TTT TGC CCC TTC TGC GAG GCT CCS TTT TYT ATC CCC ATC TGG AAC TGT TYT Het Pro TTC TGC TGC GAC CAC TGC TCC TAC ATC CCC ATC TGG AAC TTT Tyr Het Pro Tle TGG AAC TTT Tyr Het Pro Tle AAC TGC ACC CAC TTC TGC CCC ATC TGG AAC TTT TTT TTT TTT TTT TTT TTT TTT TTT TT	Asp Ser Leu Pro Asp Leu Ser Val Phe 425 GGA CGA ATT CTG CAC AAT GGC GCC TAC GIY Arg Ile Ser Trp Leu GIY Leu Arg Ser 450 CTG GCC CTC ATC CAC CAT AAC ARG ARG CCC CAC TTp Leu His Asp Clu Arg Ser 455 CTG GCC CTC ATC CAC CAT AAC ACC CAC Leu Ala Leu Ile His His Asp Clu Cys Cys Cal Axg CIV Cys Cys Cal Cac CAC Axg CIV Cys Cys Cal Cac CAC Axg CIV Cys Cys Cac CAC CAC Axg CIV Cys Cys Cac CAC CAC CAC Axg CIV Cys Cys Cac CA	Asp Ser Leu Pro Asp Leu Ser Val Phe Gln 425 GGA CGA ATT CTG CAC AAT GGC GCC TAC TCG Gly Arg 1le Leu His Asn Gly Ala Tyr Ser 435 GGC ATC AGC TGG CTG GGG CTG CGC TCA CTG CGU Ile Ser Trp Leu Gly Leu Arg Ser Leu 455 CTG GCC CTC ATC CAC CAT AAC ACC CAC CTC Leu Ala Leu Ile His His Asn Thr His Leu 465 CCC TGG GAC CAG CTC TTT CGG AAC CCG CAC Pro Trp Asp Glu 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Thr Gln Cys S50 CGA GTA CTG CAG GGG CTC CCC AGG GAG TGC GAG GGC CAC CAC CAC TGT AGG GAG CAC TGC TGC CAC AGG GAG TGC GAG GAG TGC GAG GAG CAC TGC AGG GAG TGC GAG GAG TGC GAG GAG TGC GAG GAG TGC GAG GAG CAC TGT AGG GAG GAG TGC GAG GAG GAG GAG GAG GAG GAG GAG GAG G

	TGC Cys															1968
	GTG Val															2016
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Arg	CTG Leu 690	Leu														2112
	ATG Net															2160
	AAG Lys															2208
	ATC Ile															2256
	GTG Val															2304
	GAA Glu 770															2352
	CTG Leu															2400
	CCC Pro															2448
	GGC Gly															2496
	AGC Ser															2544
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			GTG Val													2688
			ACC Thr 900													2736
			ATG Ket													2784
			CCT Pro													2832
			ACC Thr													2880
			GAA Glu													2928
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	Pro		CAG Gln			Phe					Ala					3120
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	Ala	TTT Phe				Val					Tyr					3600
		GCT Ala			Gln					Pro					Ala	3648
		AAC Asn		Tyr					Asp					Gly		3696
		AGC Ser 1235	Thr					Pro					Pro			3744
		CTG Leu					*					Ser				3792
CCC Pro 1265	*	TGT Cys				Glu					Asp					3840
CAA Gln					Pro					Gln					Cys	3888

CAG Gln	GAA Glu	CCT Pro	GTC Val 1300	Leu	AGG Arg	AAC Asn	CTT Leu	CCT Pro 1305	Ser	TGC Cys	TTG Leu	AGT Ser	TCC Ser 1310	Gln	ATG Net	3936
GCT Ala	GGA Gly	AGG Arg 1315	Gly	CCA Pro	GCC Ala	TCG Ser	TTG Leu 1320	Glu	GAG Glu	GAA Glu	CAG Gln	CAC His 1325	TGG Trp	GGA Gly	GTC Val	3984
TTT Phe	GTG Val 1330	Asp	TCT Ser	GAG Glu	GCC Ala	CTG Leu 1335	Pro	AAT Asn	GAG Glu	ACT Thr	CTA Leu 1340	Gly	TCC Ser	AGT Ser	GGA Gly	4032
TGC Cys 1345	His	AGC Ser	CCA Pro	GCT Ala	TGG Trp 1350	Pro	TTT Phe	CCT Pro	TCC Ser	AGA Arg 1355	Ser	TGG Trp	GTA Val	CTG Leu	AAA Lys 1360	4080
GCC Ala	TTA Leu	GGG Gly	AAG Lys	CTG Leu 1365	Ala	TGA *	GAG Glu	GGG Gly	AAG Lys 1370	Arg	CCC Pro	TAA *	GGG Gly	AGT Ser 1375	Val	4128
TAA *	GAA Glu	CAA Gln	AAG Lys 1380	Arg	CCC Pro	ATT Ile	CAG Gln	AGA Arg 1385	Leu	TCC Ser	CTG Leu	AAA Lys	CCT Pro 1390	Ser	ACT Thr	4176
GCC Ala	CCC Pro	CAT His 1395	Glu	GAA Glu	GGA Gly	ACA Thr	GCA Ala 1400	Het	GTG Val	TCA Ser	GTA Val	TCC Ser 140	AGG Arg	CTT Leu	TGT Cys	4224
ACA Thr	GAG Glu 141	Cys	TTT Phe	TCT Ser	GTT Val	TAG * 141	Phe	TTA Leu	CTT Leu	TTT Phe	TTG Leu 1420	Phe	TGT Cys	TTT Phe	TTT Phe	4272
	Asp	GAA Glu				Gln										4299
(2)	IN	FORM	ATIO	N FO	R SE	Q ID	NO:	2:								

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1433 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Glu Leu Ala Ala Leu Cys Arg Trp Gly Leu Leu Leu Ala Leu Leu 1 5 10 15

Pro Pro Gly Ala Ala Ser Thr Gln Val Cys Thr Gly Thr Asp Met Lys 20 25 30

Leu Arg Leu Pro Ala Ser Pro Glu Thr His Leu Asp Het Leu Arg His Leu Tyr Gln Gly Cys Gln Val Val Gln Gly Asn Leu Glu Leu Thr Tyr Leu Pro Thr Asn Ala Ser Leu Ser Phe Leu Gln Asp Ile Gln Glu Val Gln Gly Tyr Val Leu Ile Ala His Asn Gln Val Arg Gln Val Pro Leu Gln Arg Leu Arg Ile Val Arg Gly Thr Gln Leu Phe Glu Asp Asn Tyr Ala Leu Ala Val Leu Asp Asn Gly Asp Pro Leu Asn Asn Thr Thr Pro Val Thr Gly Ala Ser Pro Gly Gly Leu Arg Glu Leu Gln Leu Arg Ser Leu Thr Glu Ile Leu Lys Gly Gly Val Leu Ile Gln Arg Asn Pro Gln Leu Cys Tyr Gln Asp Thr Ile Leu Trp Lys Asp Ile Phe His Lys Asn Asn Gln Leu Ala Leu Thr Leu Ile Asp Thr Asn Arg Ser Arg Ala Cys His Pro Cys Ser Pro Met Cys Lys Gly Ser Arg Cys Trp Gly Glu Ser 195 Ser Glu Asp Cys Gln Ser Leu Thr Arg Thr Val Cys Ala Gly Gly Cys Ala Arg Cys Lys Gly Pro Leu Pro Thr Asp Cys Cys His Glu Gln Cys Ala Ala Gly Cys Thr Gly Pro Lys His Ser Asp Cys Leu Ala Cys Leu His Phe Asn His Ser Gly Ile Cys Glu Leu His Cys Pro Ala Leu Val 265 Thr Tyr Asn Thr Asp Thr Phe Glu Ser Met Pro Asn Pro Glu Gly Arg 275 Tyr Thr Phe Gly Ala Ser Cys Val Thr Ala Cys Pro Tyr Asn Tyr Leu Ser Thr Asp Val Gly Ser Cys Thr Leu Val Cys Pro Leu His Asn Gln

315

305

Glu Val Thr Ala Glu Asp Gly Thr Gln Arg Cys Glu Lys Cys Ser Lys 325 Pro Cys Ala Arg Val Cys Tyr Gly Leu Gly Het Glu His Leu Arg Glu 345 Val Arg Ala Val Thr Ser Ala Asn Ile Gln Glu Phe Ala Gly Cys Lys Lys Ile Phe Gly Ser Leu Ala Phe Leu Pro Glu Ser Phe Asp Gly Asp 370 375 Pro Ala Ser Asn Thr Ala Pro Leu Gln Pro Glu Gln Leu Gln Val Phe 395 Glu Thr Leu Glu Glu Ile Thr Gly Tyr Leu Tyr Ile Ser Ala Trp Pro 405 Asp Ser Leu Pro Asp Leu Ser Val Phe Gln Asn Leu Gln Val Ile Arg Gly Arg Ile Leu His Asn Gly Ala Tyr Ser Leu Thr Leu Gln Gly Leu Gly Ile Ser Trp Leu Gly Leu Arg Ser Leu Arg Glu Leu Gly Ser Gly Leu Ala Leu Ile His His Asn Thr His Leu Cys Phe Val His Thr Val Pro Trp Asp Gln Leu Phe Arg Asn Pro His Gln Ala Leu Leu His Thr Ala Asn Arg Pro Glu Asp Glu Cys Val Gly Glu Gly Leu Ala Cys His 505 Gln Leu Cys Ala Arg Gly His Cys Trp Gly Pro Gly Pro Thr Gln Cys Val Asn Cys Ser Gln Phe Leu Arg Gly Gln Glu Cys Val Glu Glu Cys 535 Arg Val Leu Gln Gly Leu Pro Arg Glu Tyr Val Asn Ala Arg His Cys 550 Leu Pro Cys His Pro Glu Cys Gln Pro Gln Asn Gly Ser Val Thr Cys

Phe Gly Pro Glu Ala Asp Gln Cys Val Ala Cys Ala His Tyr Lys Asp 580 585 590

Pro Pro Phe Cys Val Ala Arg Cys Pro Ser Gly Val Lys Pro Asp Leu
595 600 605

Ser	Tyr 610	Met	Pro	Ile	Trp	Lys 615	Phe	Pro	Asp	Glu	Glu 620	Gly	Ala	Cys	Gln
Pro 625	Cys	Pro	Ile	Asn	Cys 630	Thr	His	Ser	Cys	Val 635	Asp	Leu	Asp	Asp	Lys 640
Gly	Cys	Pro	Ala	Glu 645	Gln	Arg	Ala	Ser	Pro 650	Leu	Thr	Ser	Ile	Ile 655	Ser
Ala	Val	Val	Gly 660	Ile	Leu	Leu	Val	Val 665	Val	Leu	Gly	Val	Val 670	Phe	Gly
Ile	Leu	Ile 675	Lys	Arg	Arg	Gln	Gln 680	Lys	Ile	Arg	Lys	Tyr 685	Thr	Het	Arg
Arg	Leu 690	Leu	Gln	Glu	Thr	Glu 695	Leu	Val	Glu	Pro	Leu 700	Thr	Pro	Ser	Gly
Ala 705	Met	Pro	Asn	Gln	Ala 710	Gln	Met	Arg	Ile	Leu 715	Lys	Glu	Thr	Glu	Leu 720
Arg	Lys	Val	Lys	Val 725	Leu	Gly	Ser	Gly	Ala 730	Phe	Gly	Thr	Val	Tyr 735	Lys
Gly	Ile	Trp	Ile 740	Pro	Asp	Gly	Glu	Asn 745	Val	Lys	Ile	Pro	Val 750	Ala	Ile
Lys	Val	Leu 755	Arg	Glu	Asn	Thr	Ser 760	Pro	Lys	Ala	Asn	Lys 765	Glu	Ile	Leu
Asp	Glu 770	Ala	Tyr	Val	Met	Ala 775	Gly	Val	Gly	Ser	Pro 780	Tyr	Val	Ser	Arg
Leu 785	Leu	Gly	Ile	Cys	Leu 790	Thr	Ser	Thr	Val	Gln 795	Leu	Val	Thr	Gln	Leu 800
Met	Pro	Tyr	Gly	Cys 805	Leu	Leu	Asp	His	Val 810	Arg	Glu	Asn	Arg	Gly 815	Arg
Leu	Gly	Ser	Gln 820	Asp	Leu	Leu	Asn	Trp 825		Met	Gln	Ile	Ala 830	Lys	Gly
Het	Ser	Tyr 835	Leu	Glu	Asp	Val	Arg 840	Leu	Val	His	Arg	Asp 845	Leu	Ala	Ala
Arg	Asn 850	Val	Leu	Val	Lys	Ser 855	Pro	Asn	His	Val	Lys 860	Ile	Thr	Asp	Phe
Gly 865	Leu	Ala	Arg		Leu 870	Asp .	Ile	Asp	Glu	Thr. 875	Glu	Tyr	His.	.Ala.	Asp 880
Gly	Gly	Lys	Val	Pro 885	Ile	Lys	Trp	Het	Ala 890	Leu	Glu	Ser	Ile	Leu 895	Arg

- Arg Arg Phe Thr His Gln Ser Asp Val Trp Ser Tyr Gly Val Thr Val 900 905 910
- Trp Glu Leu Met Thr Phe Gly Ala Lys Pro Tyr Asp Gly Ile Pro Ala 915 920 925
- Arg Glu Ile Pro Asp Leu Leu Glu Lys Gly Glu Arg Leu Pro Gln Pro 930 935 940
- Pro Ile Cys Thr Ile Asp Val Tyr Het Ile Het Val Lys Cys Trp Het 945 950 955 960
- Ile Asp Ser Glu Cys Arg Pro Arg Phe Arg Glu Leu Val Ser Glu Phe
 965 970 975
- Ser Arg Met Ala Arg Asp Pro Gln Arg Phe Val Val Ile Gln Asn Glu 980 985 990
- Asp Leu Gly Pro Ala Ser Pro Leu Asp Ser Thr Phe Tyr Arg Ser Leu 995 1000 1005
- Leu Glu Asp Asp Asp Met Gly Asp Leu Val Asp Ala Glu Glu Tyr Leu 1010 1015 1020
- Val Pro Gln Gln Gly Phe Phe Cys Pro Asp Pro Ala Pro Gly Ala Gly 1025 1030 1035 1040
- Gly Met Val His His Arg His Arg Ser Ser Ser Thr Arg Ser Gly Gly
 1045 1050 1055
- Gly Asp Leu Thr Leu Gly Leu Glu Pro Ser Glu Glu Glu Ala Pro Arg 1060 1065 1070
- Ser Pro Leu Ala Pro Ser Glu Gly Ala Gly Ser Asp Val Phe Asp Gly 1075 1080 1085
- Asp Leu Gly Met Gly Ala Ala Lys Gly Leu Gln Ser Leu Pro Thr His 1090 1095 1100
- Asp Pro Ser Pro Leu Gln Arg Tyr Ser Glu Asp Pro Thr Val Pro Leu 1105 1110 1115 1120
- Pro Ser Glu Thr Asp Gly Tyr Val Ala Pro Leu Thr Cys Ser Pro Gln 1125 1130 1135
- Pro Glu Tyr Val Asn Gln Pro Asp Val Arg Pro Gln Pro Pro Ser Pro 1140 1145 1150
- Arg Glu Gly Pro Leu Pro Ala Ala Arg Pro Ala Gly Ala Thr Leu Glu 1155 1160 1165
- Arg Pro Lys Thr Leu Ser Pro Gly Lys Asn Gly Val Val Lys Asp Val 1170 1175 1180

Phe Ala Phe Gly Gly Ala Val Glu Asn Pro Glu Tyr Leu Thr Pro Gln 1185 1190 1195 1200

Gly Gly Ala Ala Pro Gln Pro His Pro Pro Pro Ala Phe Ser Pro Ala 1205 1210 1215

Phe Asp Asn Leu Tyr Tyr Trp Asp Gln Asp Pro Pro Glu Arg Gly Ala 1220 1225 1230

Pro Pro Ser Thr Phe Lys Gly Thr Pro Thr Ala Glu Asn Pro Glu Tyr 1235 1240 1245

Leu Gly Leu Asp Val Pro Val * Thr Arg Arg Pro Ser Pro Gln Lys 1250 1255 1260

Pro * Cys Val Leu Arg Glu Gln Gly Arg Pro Asp Phe Cys Trp His 1265 1270 1275 1280

Gln Glu Val Gly Gly Pro Ser Asp His Phe Gln Gly Asn Leu Pro Cys 1285 1290 1295

Gln Glu Pro Val Leu Arg Asn Leu Pro Ser Cys Leu Ser Ser Gln Met 1300 1305 1310

Ala Gly Arg Gly Pro Ala Ser Leu Glu Glu Glu Gln His Trp Gly Val 1315 1320 1325

Phe Val Asp Ser Glu Ala Leu Pro Asn Glu Thr Leu Gly Ser Ser Gly 1330 1335 1340

Cys His Ser Pro Ala Trp Pro Phe Pro Ser Arg Ser Trp Val Leu Lys 1345 1350 1355 1360

Ala Leu Gly Lys Leu Ala * Glu Gly Lys Arg Pro * Gly Ser Val 1365 1370 1375

* Glu Gln Lys Arg Pro Ile Gln Arg Leu Ser Leu Lys Pro Ser Thr 1380 1385 1390

Ala Pro His Glu Glu Gly Thr Ala Met Val Ser Val Ser Arg Leu Cys 1395 1400 1405

Thr Glu Cys Phe Ser Val * Phe Leu Leu Phe Leu Phe Cys Phe Phe 1410 1415 1420

Lys Asp Glu Ile Lys Thr Gln Gly Glu 1425 1430

(2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 739 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

- (A) NAME/KEY: CDS

 - (B) LOCATION: 1..739
 (D) OTHER INFORMATION: /note= "product = "520C9sFv/ amino acid info: 520C9sFv protein""

SEQUENCE DESCRIPTION: SEQ ID NO:3: (xi)

	Ile		Gln	TCT Ser						48
				AAG Lys						96
				CAG Gln						144
				ACT Thr 55						192
				TCT Ser						240
				AGA Arg						288
				GCT Ala						336
				AGC Ser						384
Ser				ATC Ile 135						432
				AGA Arg						480
				ACC Thr						528

- 61 -

		CGC Arg														576
		TTC Phe 195														624
		CTT Leu														672
		TTT Phe														720
	GCT Ala	GAT Asp	TAA *	TCT Ser 245	_	G										739
(2)	IN	FORMA	TION	v FOF	R SEC) ID	NO:4	:								
		(i)	(<i>E</i>	A) I	ICE (LENGT TYPE:	TH: 2 ami	246 a	umino acid		ids						
		(ii)	MC	DLECU	ILE 1	TYPE:	pro	teir	1							
		(xi)	SE	EQUEN	ICE I	ESCF	RIPT	ON:	SEQ	ID N	10:4:	:				
Glu 1	Ile	Gln	Leu	Val 5	Gln	Ser	Gly	Pro	Glu 10	Leu	Lys	Lys	Pro	Gly 15	Glu	
Thr	Val	Lys	Ile 20	Ser	Cys	Lys	Ala	Ser 25	Gly	Tyr	Thr	Phe	Ala 30	Asn	Tyr	
Gly	Met	Asn 35	Trp	Met	Lys	Gln	Ala 40	Pro	Gly	Lys	Gly	Leu 45	Lys	Trp	Met	
Gly	Trp 50	Ile	Asn	Thr	Tyr	Thr 55	Gly	Gln	Ser	Thr	Tyr 60	Ala	Asp	Asp	Phe	
Lys 65	Glu	Arg	Phe	Ala	Phe 70	Ser	Leu	Glu	Thr	Ser 75	Ala	Thr	Thr	Ala	His 80	
Leu	Gln	Ile	Asn	Asn 85		Arg	Asn	Glu	Asp 90	Ser	Ala	Thr	Tyr	Phe 95	Cys	
Ala	Arg	Arg	Phe 100	Gly	Phe	Ala	Tyr	Trp 105	Gly	Gln	Gly	Thr	Leu 110	Val	Ser	
Val	Ser	Ala	Ser	Ile	Ser	Ser	Ser	Ser	Gly	Ser	Ser	Ser	Ser	Gly	Ser	

Ser Ser Ser Gly Ser Asp Ile Gln Het Thr Gln Ser Pro Ser Ser Leu 135 Ser Ala Ser Leu Gly Glu Arg Val Ser Leu Thr Cys Arg Ala Ser Gln 150 155 Asp Ile Gly Asn Ser Leu Thr Trp Leu Gln Gln Glu Pro Asp Gly Thr Ile Lys Arg Leu Ile Tyr Ala Thr Ser Ser Leu Asp Ser Gly Val Pro 185 Lys Arg Phe Ser Gly Ser Arg Ser Gly Ser Asp Tyr Ser Leu Thr Ile 195 200 Ser Ser Leu Glu Ser Glu Asp Phe Val Val Tyr Tyr Cys Leu Gln Tyr 215 Ala Ile Phe Pro Tyr Thr Phe Gly Gly Gly Thr Asn Leu Glu Ile Lys Arg Ala Asp Ser Ala 245 INFORMATION FOR SEQ ID NO:5: DELETED ACCORDING TO PRELIMINARY AMENDMENT (2) INFORMATION FOR SEQ ID NO:6: DELETED ACCORDING TO PRELIMINARY AMENDMENT (2) INFORMATION FOR SEQ IS NO:7: SEQUENCE CHARACTERISTICS: LENGTH: 807 base pairs (A) (B) TYPE: nucleic acid (C) STRANDEDNESS: single TOPOLOGY: linear (D) MOLECULE TYPE: DNA (genomic) (ii) (ix) FEATURE: (A) NAME/KEY: CDS LOCATION: 1..807 (B) (D) OTHER INFORMATION: /note= "product = "Ricin-A chain gene/ amino acid info: Ricin-A chain protein"" SEQUENCE DESCRIPTION: SEQ ID NO:7: (xi) ATG ATA TTC CCC AAA CAA TAC CCA ATT ATA AAC TTT ACC ACA GCG GGT 48 Met Ile Phe Pro Lys Gln Tyr Pro Ile Ile Asn Phe Thr Thr Ala Gly 10 GCC ACT GTG CAA AGC TAC ACA AAC TTT ATC AGA GCT GTT CGC GGT CGT 96 Ala Thr Val Gln Ser Tyr Thr Asn Phe Ile Arg Ala Val Arg Gly Arg

25

20

ACA Thr									144
GTT Val 50									192
CAT His									240
GTG Val									288
AAT Asn									336
AAT Asn									384
CTT Leu 130									432
GAG Glu							_		480
CTT Leu									528
GAA Glu									576
AGG Arg									624
AAT Asn 210									672
GCC Ala									720
AGT Ser									768

GTG TAT AGA TGC GCA CCT CCA CCA TCG TCA CAG TTT TAA
Val Tyr Arg Cys Ala Pro Pro Pro Ser Ser Gln Phe
260
265

807

(2) INFORMATION FOR SEQ ID NO:8:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 268 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Met Ile Phe Pro Lys Gln Tyr Pro Ile Ile Asn Phe Thr Thr Ala Gly
1 5 10 15

Ala Thr Val Gln Ser Tyr Thr Asn Phe Ile Arg Ala Val Arg Gly Arg
20 25 30

Leu Thr Thr Gly Ala Asp Val Arg His Glu Ile Pro Val Leu Pro Asn 35 40 45

Arg Val Gly Leu Pro Ile Asn Gln Arg Phe Ile Leu Val Glu Leu Ser 50 55 60

Asn His Ala Glu Leu Ser Val Thr Leu Ala Leu Asp Val Thr Asn Ala 65 70 .75 80

Tyr Val Val Gly Tyr Arg Ala Gly Asn Ser Ala Tyr Phe Phe His Pro 85 90 95

Asp Asn Gln Glu Asp Ala Glu Ala Ile Thr His Leu Phe Thr Asp Val

Gln Asn Arg Tyr Thr Phe Ala Phe Gly Gly Asn Tyr Asp Arg Leu Glu 115 120 125

Gln Leu Ala Gly Asn Leu Arg Glu Asn Ile Glu Leu Gly Asn Gly Pro 130 135 140

Leu Glu Glu Ala Ile Ser Ala Leu Tyr Tyr Tyr Ser Thr Gly Gly Thr 145 150 155 160

Gln Leu Pro Thr Leu Ala Arg Ser Phe Ile Ile Cys Ile Gln Het Ile 165 170 175

Ser Glu Ala Ala Arg Phe Gln Tyr Ile Glu Gly Glu Het Arg Thr Arg 180 185 190

Ile Arg Tyr Asn Arg Arg Ser Ala Pro Asp Pro Ser Val Ile Thr Leu 195 200 205

Glu	Asn 210	Ser	Trp	Gly	Arg	Leu 215	Ser	Thr	Ala	Ile	Gln 220	Glu	Ser	Asn	Gln	
Gly 225	Ala	Phe	Ala	Ser	Pro 230	Ile	Gln	Leu	Gln	Arg 235	Arg	Asn	Gly	Ser	Lys 240	
Phe	Ser	Val	Tyr	Asp 245	Val	Ser	Ile	Leu	Ile 250	Pro	Ile	Ile	Ala	Leu 255	Het	
Val	Tyr	Arg	Cys 260	Ala	Pro	Pro	Pro	Ser 265	Ser	Gln	Phe					
(2)	(2) INFORMATION FOR SEQ ID NO:9:															
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1605 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear																
		(ii)) M(LECU	ILE 7	YPE:	: DNA	A (ge	enomi	ic)						
		(ix)	(<i>I</i>	3) 1	AME,	CION	1.	. 160	5 N. /*		****		·• ·-	"G_1	277911	
			(1	0) (THE	(TM	UNI	ATTU	M: \1	iote=	= "p	couuc	.L =	G-1	FIT""	
		(xi)	,	EQUE									JL =	G		
AAG Lys 1	CTT Leu	ATG	SI ATA	Ť	CCC	DESCI	RIPT:	ION:	SEQ CCA	ID I	:0:9 ATA	: AAC	TTT	ACC	ACA	48
Lys 1	Leu	ATG Met	ATA Ile	EQUEN TTC Phe	CCC Pro	AAA Lys AGC	CAA Gln TAC	ION: TAC Tyr	SEQ CCA Pro 10 AAC Asn	ID PATT Ile	ATA Ile	AAC Asn	TTT Phe GCT	ACC Thr 15	ACA Thr	48 96
Lys 1 GCG Ala	GGT Gly	ATG Met GCC Ala	ATA Ile ACT Thr 20	TTC Phe 5	CCC Pro CAA Gln	AAA Lys AGC Ser	CAA Gln TAC Tyr	TAC Tyr ACA Thr 25	SEQ CCA Pro 10 AAC Asn	ID 1 ATT Ile TTT Phe	ATA Ile ATC Ile	AAC Asn AGA Arg	TTT Phe GCT Ala 30	ACC Thr 15 GTT Val	ACA Thr CGC Arg	
Lys 1 GCG Ala GGT Gly	GGT Gly CGT Arg	ATG Met GCC Ala TTA Leu 35	ATA Ile ACT Thr 20 ACA Thr	TTC Phe 5 GTG Val	CCC Pro CAA Gln GGA Gly	AAA Lys AGC Ser GCT Ala	CAA Gln TAC Tyr GAT Asp 40	TAC Tyr ACA Thr 25 GTG Val	SEQ CCA Pro 10 AAC Asn AGA Arg	ID PATT Ile TTT Phe CAT His	ATA Ile ATC Ile GAA Glu	AAC Asn AGA Arg ATA Ile 45 ATT Ile	TTT Phe GCT Ala 30 CCA Pro	ACC Thr 15 GTT Val GTG Val	ACA Thr CGC Arg TTG Leu	96
Lys 1 GCG Ala GGT Gly CCA Pro	GGT Gly CGT Arg AAC Asn 50	ATG Met GCC Ala TTA Leu 35 AGA Arg	ATA Ile ACT Thr 20 ACA Thr GTT Val	TTC Phe 5 GTG Val ACT Thr	CCC Pro CAA Gln GGA Gly TTG Leu	AAA Lys AGC Ser GCT Ala CCT Pro 55	CAA Gln TAC Tyr GAT Asp 40 ATA Ile	TAC Tyr ACA Thr 25 GTG Val AAC Asn	SEQ CCA Pro 10 AAC Asn AGA Arg CAA Gln	ID PATT Ile TTT Phe CAT His CGG Arg	ATA Ile ATC Ile GAA Glu TTT Phe 60 GCG Ala	AAC Asn AGA Arg ATA Ile 45 ATT Ile CTG	TTT Phe GCT Ala 30 CCA Pro TTA Leu	ACC Thr 15 GTT Val GTG Val GTT Val	ACA Thr CGC Arg TTG Leu GAA Glu	96 144

CAT His	CCT Pro	GAC Asp	AAT Asn 100	CAG Gln	GAA Glu	GAT Asp	GCA Ala	GAA Glu 105	GCA Ala	ATC Ile	ACT Thr	CAT His	CTT Leu 110	TTC Phe	ACT Thr	3	336
GAT Asp	GTT Val	CAA Gln 115	Asn	CGA Arg	TAT Tyr	ACA Thr	TTC Phe 120	GCC Ala	TTT Phe	GGT Gly	GGT Gly	AAT Asn 125	TAT Tyr	GAT Asp	AGA Arg		384
											ATC Ile 140				AAT Asn	. 4	432
GGT Gly 145	CCA Pro	CTA Leu	GAG Glu	GAG Glu	GCT Ala 150	ATC Ile	TCA Ser	GCG Ala	CTT Leu	TAT Tyr 155	TAT Tyr	TAC Tyr	AGT Ser	ACT Thr	GGT Gly 160		480
GGC Gly	ACT Thr	CAG Gln	CTT Leu	CCA Pro 165	ACT Thr	CTG Leu	GCT Ala	CGT Arg	TCC Ser 170	TTT Phe	ATA Ile	ATT Ile	TGC Cys	ATC Ile 175	CAA Gln	5	528
ATG Met	ATT Ile	TCA Ser	GAA Glu 180	GCA Ala	GCA Ala	AGA Arg	TTC Phe	CAA Gln 185	TAT Tyr	ATT Ile	GAG Glu	GGA Gly	GAA Glu 190	ATG Met	CGC Arg		576
											GAT Asp					6	524
ACA Thr	CTT Leu 210	GAG Glu	AAT Asn	AGT Ser	TGG Trp	GGG Gly 215	AGA Arg	CTT Leu	TCC Ser	ACT Thr	GCA Ala 220	ATT Ile	CAA Gln	GAG Glu	TCT Ser	. 6	572
											CAA Gln					7	720
TCC Ser	AAA Lys	TTC Phe	AGT Ser	GTG Val 245	TAC Tyr	GAT Asp	GTG Val	AGT Ser	ATA Ile 250	TTA Leu	ATC Ile	CCT Pro	ATC Ile	ATA Ile 255	GCT Ala	7	768
CTC Leu	ATG Het	GTG Val	TAT Tyr 260	AGA Arg	TGC Cys	GCA Ala	CCT Pro	CCA Pro 265	CCA Pro	TCG Ser	TCA Ser	CAG Gln	TTT Phe 270	TCT Ser	CTT Leu	8	316
CTT Leu	ATA Ile	AGG Arg 275	CCA Pro	GTG Val	GTA Val	CCA Pro	AAT Asn 280	TTT Phe	AAT Asn	GCT Ala	GAT Asp	GTT Val 285	TGT Cys	ATG Met	GAT Asp	. 8	364
CCT Pro	GAG Glu 290	ATC Ile	CAA Gln	TTG Leu	GTG Val	CAG Gln 295	TCT Ser	GGA Gly	CCT Pro	GAG Glu	CTG Leu 300	Lys	AAG Lys	CCT Pro	GGA Gly	9	91-2

GAG Glu 305	ACA Thr	GTC Val	AAG Lys	ATC Ile	TCC Ser 310	TGC Cys	AAG Lys	GCT Ala	TCT Ser	GGA Gly 315	TAT Tyr	ACC Thr	TTC Phe	GCA Ala	AAC Asn 320	960
TAT Tyr	GGA Gly	ATG Het	AAC Asn	TGG Trp 325	ATG Met	AAG Lys	CAG Gln	GCT Ala	CCA Pro 330	GGA Gly	AAG Lys	GGT Gly	TTA Leu	AAG Lys 335	TGG Trp	1008
ATG Met	GGC Gly	TGG Trp	ATA Ile 340	AAC Asn	ACC Thr	TAC Tyr	ACT Thr	GGA Gly 345	CAG Gln	TCA Ser	ACA Thr	TAT Tyr	GCT Ala 350	GAT Asp	GAC Asp	1056
TTC Phe	AAG Lys	GAA Glu 355	CGG Arg	TTT Phe	GCC Ala	TTC Phe	TCT Ser 360	TTG Leu	GAA Glu	ACC Thr	TCT Ser	GCC Ala 365	ACC Thr	ACT Thr	GCC Ala	1104
CAT His	TTG Leu 370	CAG Gln	ATC Ile	AAC Asn	AÁĈ Asn	ČŤĊ Leu 375	AĞA Arg	AAT Asn	GAG Glu	GAC Asp	TCG Ser 380	GCC Ala	ACA Thr	TAT Tyr	TTC Phe	1152
TGT Cys 385	GCA Ala	AGA Arg	CGA Arg	TTT Phe	GGG Gly 390	TTT Phe	GCT Ala	TAC Tyr	TGG Trp	GGC Gly 395	CAA Gln	GGG Gly	ACT Thr	CTG Leu	GTC Val 400	1200
AGT Ser	GTC Val	TCT Ser	GCA Ala	TCG Ser 405	ATA Ile	TCG Ser	AGC Ser	TCT Ser	GGT Gly 410	GGC Gly	GGT Gly	GGC Gly	TCG Ser	GGC Gly 415	GGT Gly	1248
GGT Gly	GGG Gly	TCG Ser	GGT Gly 420	GGC Gly	GGC Gly	GGA Gly	TCG Ser	GAT Asp 425	ATC Ile	CAG Gln	ATG Met	ACC Thr	CAG Gln 430	TCT Ser	CCA Pro	1296
TCC Ser	TCC Ser	TTA Leu 435	TCT Ser	GCC Ala	TCT Ser	CTG Leu	GGA Gly 440	GAA Glu	AGA Arg	GTC Val	AGT Ser	CTC Leu 445	ACT Thr	TGT Cys	CGG Arg	1344
GCA Ala	AGT Ser 450	CAG Gln	GAC Asp	ATT Ile	GGT Gly	AAT Asn 455	AGC Ser	TTA Leu	ACC Thr	TGG Trp	CTT Leu 460	TCA Ser	CAG Gln	GAA Glu	CCA Pro	1392
GAT Asp 465	GGA Gly	ACT Thr	ATT Ile	AAA Lys	CGC Arg 470	CTG Leu	ATC Ile	TAC Tyr	GCC Ala	ACA Thr 475	TCC Ser	AGT Ser	TTA Leu	GAT Asp	TCT Ser 480	1440
GGT Gly	GTC Val	CCC Pro	AAA Lys	AGG Arg 485	TTC Phe	AGT Ser	GGC Gly	AGT Ser	CGG Arg 490	TCT Ser	GGG Gly	TCA Ser	GAT Asp	TAT Tyr 495	TCT Ser	1488
CTC Leu	ACC Thr	ATC Ile	AGT Ser 500	AGC Ser	CTT Leu	GAG Glu	TCT Ser	GAA Glu 505	GAT Asp	TTT Phe	GTA Val	GTC Val	TAT Tyr 510	TAC	TGT Cys	1536
CTA Leu	CAA Gln	TAT Tyr 515	GCT Ala	ATT	TTT Phe	CCG Pro	TAC Tyr 520	ACG Thr	TTC Phe	GGA Gly	GGG Gly	GGG Gly 525	ACC Thr	AAC Asn	CTG Leu	1584

GAA ATA AAA CGG GCT GAT TAA Glu Ile Lys Arg Ala Asp 530 535 1605

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 534 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Lys Leu Met Ile Phe Pro Lys Gln Tyr Pro Ile Ile Asn Phe Thr Thr

Ala Gly Ala Thr Val Gln Ser Tyr Thr Asn Phe Ile Arg Ala Val Arg 20 25 30

Gly Arg Leu Thr Thr Gly Ala Asp Val Arg His Glu Ile Pro Val Leu
35 40 45

Pro Asn Arg Val Gly Leu Pro Ile Asn Gln Arg Phe Ile Leu Val Glu 50 55 60

Leu Ser Asn His Ala Glu Leu Ser Val Thr Leu Ala Leu Asp Val Thr 65 70 75 80

Asn Ala Tyr Val Val Gly Tyr Arg Ala Gly Asn Ser Ala Tyr Phe Phe 85 90 95

His Pro Asp Asn Gln Glu Asp Ala Glu Ala Ile Thr His Leu Phe Thr

Asp Val Gln Asn Arg Tyr Thr Phe Ala Phe Gly Gly Asn Tyr Asp Arg

Leu Glu Gln Leu Ala Gly Asn Leu Arg Glu Asn Ile Glu Leu Gly Asn 130 135 140

Gly Pro Leu Glu Glu Ala Ile Ser Ala Leu Tyr Tyr Tyr Ser Thr Gly 145 150 155 160

Gly Thr Gln Leu Pro Thr Leu Ala Arg Ser Phe Ile Ile Cys Ile Gln 165 170 175

Met Ile Ser Glu Ala Ala Arg Phe Gln Tyr Ile Glu Gly Glu Met Arg 180 185 190

Thr Arg Ile Arg Tyr Asn Arg Arg Ser Ala Pro Asp Pro Ser Val Ile 195 200 205

Thr Leu Glu Asn Ser Trp Gly Arg Leu Ser Thr Ala Ile Gln Glu Ser Asn Gln Gly Ala Phe Ala Ser Pro Ile Gln Leu Gln Arg Arg Asn Gly 230 Ser Lys Phe Ser Val Tyr Asp Val Ser Ile Leu Ile Pro Ile Ile Ala 250 Leu Met Val Tyr Arg Cys Ala Pro Pro Pro Ser Ser Gln Phe Ser Leu 265 Leu Ile Arg Pro Val Val Pro Asn Phe Asn Ala Asp Val Cys Het Asp 280 Pro Glu Ile Gln Leu Val Gln Ser Gly Pro Glu Leu Lys Lys Pro Gly Glu Thr Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Ala Asn Tyr Gly Met Asn Trp Met Lys Gln Ala Pro Gly Lys Gly Leu Lys Trp Met Gly Trp Ile Asn Thr Tyr Thr Gly Gln Ser Thr Tyr Ala Asp Asp Phe Lys Glu Arg Phe Ala Phe Ser Leu Glu Thr Ser Ala Thr Thr Ala His Leu Gln Ile Asn Asn Leu Arg Asn Glu Asp Ser Ala Thr Tyr Phe Cys Ala Arg Arg Phe Gly Phe Ala Tyr Trp Gly Gln Gly Thr Leu Val 395 400 Ser Val Ser Ala Ser Ile Ser Ser Ser Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser Asp Ile Gln Met Thr Gln Ser Pro Ser Ser Leu Ser Ala Ser Leu Gly Glu Arg Val Ser Leu Thr Cys Arg Ala Ser Gln Asp Ile Gly Asn Ser Leu Thr Trp Leu Ser Gln Glu Pro 455 Asp Gly Thr Ile Lys Arg Leu Ile Tyr Ala Thr Ser Ser Leu Asp Ser 465 Gly Val Pro Lys Arg Phe Ser Gly Ser Arg Ser Gly Ser Asp Tyr Ser 485 490

Leu Thr Ile Ser Ser Leu Glu Ser Glu Asp Phe Val Val Tyr Tyr Cys 500 505 510

Leu Gln Tyr Ala Ile Phe Pro Tyr Thr Phe Gly Gly Gly Thr Asn Leu 515 520 525

Glu Ile Lys Arg Ala Asp 530

- (2) INFORMATION FOR SEQ ID NO:11:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 1..45
 - (D) OTHER INFORMATION: /note= "product = "new linker/ info: new linker""
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

TCG AGC TCC TCC GGA TCT TCA TCT AGC GGT TCC AGC TCG AGT GGA Ser Ser Ser Ser Gly Ser Ser Ser Gly Ser Ser Ser Gly 1 5 10 15 45

- (2) INFORMATION FOR SEQ ID NO:12:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 15 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ser Ser Ser Ser Gly Ser Ser Ser Ser Gly Ser Ser Ser Ser Gly
1 5 10 15

- (2) INFORMATION FOR SEQ ID NO:13:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 45 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

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(ii) MOLECULE TYPE: DNA (genomic)

(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 145 (D) OTHER INFORMATION: /note= "product = "old linker/ protein info: old linker""
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:13:
	GA TCT GGA GGA GGA TCT GGA GGA GGA GGA TCT ly Ser Gly Gly Gly Ser Gly Gly Gly Ser 5 10 15
(2) INFORMAT	ION FOR SEQ ID NO:14:
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 15 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear
(ii)	MOLECULE TYPE: protein
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:14:
Gly Gly Gly G	ly Ser Gly Gly Gly Gly Ser Gly Gly Gly Ser 5 10 15
(2) INFORMATI	ION FOR SEQ ID NO:15:
(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 2001 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear
(ii)	MOLECULE TYPE: DNA (genomic)
(ix)	FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 12001 (D) OTHER INFORMATION: /note= "product = "741sFv-PE40""
(xí)	SEQUENCE DESCRIPTION: SEQ ID NO:15:
	CC CAA TTG GTG CAG TCT GGA CCT GAG CTG AAG AAG CCT Le Gln Leu Val Gln Ser Gly Pro Glu Leu Lys Lys Pro 5 10 15
Gly Glu Thr Va	CC AAG ATC TCC TGC AAG GCT TCT GGG TAT ACC TTC ACA 11 Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr 25 30

	Het		GTG Val							144
			ACC Thr 55					GAA Glu		192
			GCC Ala							240
			AAC Asn							288
			ATT Ile							33 6
			GTC Val							384
			AGC Ser 135							432
			GGA Gly							480
			GCT Ala							528
			ATT Ile							576
			GGC Gly							624
			GCT Ala 215						·	672
			TAC Tyr							720
			GCA Ala							768

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		GGC Gly						816
		CTG Leu						864
		GAG Glu						912
		GCG Ala 310						960
		GCC Ala					Glu	1008
		CCG Pro						1056
		CGC Arg				Asn		1104
		GCC Ala						1152
		GGC Gly 390						1200
		GGC Gly						1248
		GGC Gly						1296
		CTG Leu						1344
 		GAA Glu						1392
		GAC Asp 470						1440

	GGC Gly																1488
	GCA Ala			Arg													1536
	CGC Arg																1584
GCG Ala	CCG Pro 530	GAG Glu	GCG Ala	GCG Ala	GGC Gly	GAG Glu 535	GTC Val	GAA Glu	CGG Arg	CTG Leu	ATC Ile 540	GGC Gly	CAT His	CCG Pro	CTG Leu		1632
	CTG Leu																1680
CTG Leu	GAG Glu	ACC Thr	ATT Ile	CTC Leu 565	GGC Gly	TGG Trp	CCG Pro	CTG Leu	GCC Ala 570	GAG Glu	CGC Arg	ACC Thr	GTG Val	GTG Val 575	ATT Ile		1728
	TCG Ser																1776
	TCC Ser																1824
	GCC Ala 610															·	1872
	CGA Arg														CTG Leu 640		1920
GCC Ala	ATA Ile	CAT His	CAG Gln	GTT Val 645	TTC Phe	CTG Leu	ATG Het	CCA Pro	GCC Ala 650	CAA Gln	TCG Ser	AAT Asn	ATG Met	AAT Asn 655	TGA *		1968
	TCT Ser																2001

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 667 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Asp Pro Glu Ile Gln Leu Val Gln Ser Gly Pro Glu Leu Lys Lys Pro Gly Glu Thr Val Lys Ile Ser Cys Lys Ala Ser Gly Tyr Thr Phe Thr Asn Tyr Gly Het Asn Trp Val Lys Gln Ala Pro Gly Lys Gly Leu Lys Trp Met Gly Trp Ile Asn Thr Asn Thr Gly Glu Pro Thr Tyr Ala Glu Glu Phe Lys Gly Arg Phe Ala Phe Ser Leu Glu Thr Ser Ala Ser Thr Ala Tyr Leu Gln Ile Asn Asn Leu Lys Asn Glu Asp Thr Ala Thr Tyr Phe Cys Gly Arg Gln Phe Ile Thr Tyr Gly Gly Phe Ala Asn Trp Gly Gln Gly Thr Leu Val Thr Val Ser Ala Ser Ser Ser Ser Gly Ser Ser 120 Ser Ser Gly Ser Ser Ser Ser Asp Ile Val Met Thr Gln Ser Pro Lys 135 Phe Met Ser Thr Ser Val Gly Asp Arg Val Ser Ile Ser Cys Lys Ala 145 Ser Gln Asp Val Ser Thr Ala Val Ala Trp Tyr Gln Gln Lys Pro Gly Gln Ser Pro Lys Leu Leu Ile Tyr Trp Thr Ser Thr Arg His Thr Gly 180 Val Pro Asp Pro Phe Thr Gly Ser Gly Ser Gly Thr Asp Tyr Thr Leu Thr Ile Ser Ser Val Gln Ala Glu Asp Leu Ala Leu His Tyr Cys Gln 215 Gln His Tyr Arg Val Ala Tyr Thr Phe Gly Arg Gly Thr Lys Leu Glu 235 230 Ile Lys Arg Ala Asp Ala Ala Pro Thr Val Ser Ile Phe Pro Pro Ser 250 Ser Glu Gln Phe Glu Gly Gly Ser Leu Ala Ala Leu Asn Ala His Gln 260 265

Ala Cys His Leu Pro Leu Glu Thr Phe Thr Arg His Arg Gln Pro Arg 280 Gly Trp Glu Gln Leu Glu Gln Cys Gly Tyr Pro Val Gln Arg Leu Val 300 Ala Leu Tyr Leu Ala Ala Arg Leu Ser Trp Asn Gln Val Asp Gln Val Ile Arg Asn Ala Leu Ala Ser Pro Gly Ser Gly Gly Asp Leu Gly Glu 330 Ala Ile Arg Glu Gln Pro Glu Gln Ala Arg Leu Ala Leu Thr Leu Ala 340 Ala Ala Glu Ser Glu Arg Phe Val Arg Gln Gly Thr Gly Asn Asp Glu 360 Ala Gly Ala Ala Asn Ala Asp Val Val Ser Leu Thr Cys Pro Val Ala Ala Gly Glu Cys Ala Gly Pro Ala Asp Ser Gly Asp Ala Leu Leu Glu Arg Asn Tyr Pro Thr Gly Ala Glu Phe Leu Gly Asp Gly Gly Asp Val Ser Phe Ser Asn Arg Gly Thr Gln Asn Trp Thr Val Glu Arg Leu Leu Gln Ala His Arg Gln Leu Glu Glu Arg Gly Tyr Val Phe Val Gly Tyr His Gly Thr Phe Leu Glu Ala Ala Gln Ser Ile Val Phe Gly Gly Val 455 450 Arg Ala Arg Ser Gln Asp Leu Asp Ala Ile Trp Arg Gly Phe Tyr Ile 470 Ala Gly Asp Pro Ala Leu Ala Tyr Gly Tyr Ala Gln Asp Gln Glu Pro Asp Ala Arg Gly Arg Ile Arg Asn Gly Ala Leu Leu Arg Val Tyr Val Pro Arg Ser Ser Leu Pro Gly Phe Tyr Arg Thr Ser Leu Thr Leu Ala 515 Ala Pro Glu Ala Ala Gly Glu Val Glu Arg Leu Ile Gly His Pro Leu 530 Pro Leu Arg Leu Asp Ala Ile Thr Gly Pro Glu Glu Glu Gly Gly Arg Leu Glu Thr Ile Leu Gly Trp Pro Leu Ala Glu Arg Thr Val Val Ile 565 570 575

Pro Ser Ala Ile Pro Thr Asp Pro Arg Asn Val Gly Gly Asp Leu Asp 580 585 590

Pro Ser Ser Ile Pro Asp Lys Glu Gln Ala Ile Ser Ala Leu Pro Asp 595 600 605

Tyr Ala Ser Gln Pro Gly Lys Pro Pro Arg Glu Asp Leu Lys * Leu 610 615 620

Pro Arg Pro Ala Gly Ser Leu Arg Arg Ser Arg Pro Ser Arg Gly Leu 625 630 635 640

Ala Ile His Gln Val Phe Leu Met Pro Ala Gln Ser Asn Met Asn * 645 650 655

Ser Ser Arg Val Asp Leu Gln Ala Cys Lys Leu 660 665

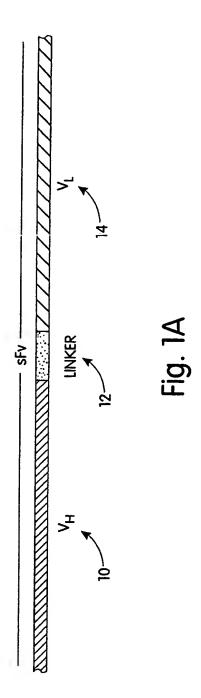
CLAIMS

- 1 1. A single-chain Fv (sFv) polypeptide defining a
- 2 binding site which exhibits the immunological binding
- 3 properties of an immunoglobulin molecule which binds
- 4 c-erbB-2 or a c-erbB-2-related tumor antigen, said sFv
- 5 comprising at least two polypeptide domains connected
- 6 by a polypeptide linker spanning the distance between
- 7 the C-terminus of one domain and the N-terminus of the
- 8 other, the amino acid sequence of each of said
- 9 polypeptide domains comprising a set of complementarity
- 10 determining regions (CDRs) interposed between a set of
- 11 framework regions (FRs), said CDRs conferring
- 12 immunological binding to said c-erbB-2 or c-erbB-2-
- 13 related tumor antigen.
 - 1 2. The single-chain Fv polypeptide of claim 1
 - 2 wherein said CDRs are substantially homologous with the
 - 3 CDRs of the c-erbB-2-binding immunoglobulin molecules
 - 4 selected from the group consisting of 520C9, 741F8, and
 - 5 454C11 monoclonal antibodies.
 - 1 3. The single-chain Fv polypeptide of claim 2
 - 2 wherein the amino acid sequence of each of said sFv
 - 3 CDRs and each of said FRs are substantially homologous
 - 4 with the amino acid sequence of CDRs and FRs of the
 - 5 variable region of 520C9 antibody.
 - 1 4. The single-chain Fv polypeptide of claim 1
 - 2 wherein said polypeptide linker comprises the amino
 - 3 acid sequence as set forth in the Sequence Listing as
 - 4 amino acid residue numbers 118 through 133 in SEQ ID
 - 5 NO:4.

- 1 5. The single-chain Fv polypeptide of claim 1
- 2 wherein said polypeptide linker comprises an amino acid
- 3 sequence selected from the group of sequences set forth
- 4 as amino acid residues 116-135 in SEQ ID NO:6, or 122-
- 5 135 in SEQ. ID NO:15 and the amino acid sequences set
- 6 forth in SEQ ID NO: 12 and SEQ ID NO: 14.
- 1 6. The single-chain Fv polypeptide of claim 1
- 2 further comprising a remotely detectable moiety bound
- 3 thereto to permit imaging of a cell bearing said
- 4 c-erbB-2-related tumor antigen.
- 1 7. The single-chain Fv polypeptide of claim 6
- 2 wherein said remotely detectable moiety comprises a
- 3 radioactive atom.
- 1 8. The single-chain Fv polypeptide of claim 1
- 2 further comprising, linked to the N or C terminus of
- 3 said linked domains, a third polypeptide domain
- 4 comprising an amino acid sequence defining CDRs
- 5 interposed between FRs and defining a second
- 6 immunologically active site.
- 1 9. The single-chain Fv polypeptide of claim 8,
- 2 further comprising a fourth polypeptide domain, wherein
- 3 said third and fourth polypeptide domains together
- 4 comprise a second site which immunologically binds a
- 5 c-erbB-2-related tumor antigen.
- 1 10. The single-chain Fv polypeptide of claim 1 or 7
- 2 further comprising a toxin linked to the N or C
- 3 terminus of said linked domain.

- 1 11. The single-chain Fv polypeptide of claim 10
- 2 wherein said toxin comprises a toxic portion selected
- 3 from the group: Pseudomonas exotoxin, ricin, ricin A
- 4 chain, phytolaccin and diphtheria toxin.
- 1 12. The single-chain Fv polypeptide of claim 10
- 2 wherein said toxin comprises at least a portion of the
- 3 ricin A chain.
- 1 13. A DNA sequence encoding the polypeptide chain of
- 2 claim 1.
- 1 14. A method of producing a single chain polypeptide
- 2 having specificity for a c-erbB-2-related tumor
- 3 antigen, said method comprising the steps of:
- 4 (a) transfecting the DNA of claim 13 into a
- 5 host cell to produce a transformant; and
- 6 (b) culturing said transformant to produce
- 7 said single-chain polypeptide.
- 1 15. A method of imaging a tumor expressing a
- 2 c-erbB-2-related antigen, said method comprising the
- 3 steps of:
- 4 (a) providing an imaging agent comprising the
- 5 polypeptide of claim 7;
- 6 (b) administering to a mammal harboring said
- 7 tumor an amount of said imaging agent together with a
- 8 physiologically-acceptable carrier sufficient to permit
- 9 extracorporeal detection of said tumor after allowing
- 10 said agent to bind to said tumor; and
- 11 (c) detecting the location of said remotely
- 12 detectable moiety in said subject to obtain an image of
- 13 said tumor.

- 1 16. A host cell transfected with a DNA of claim 13.
- 1 17. A method of inhibiting in vivo growth of a tumor
- 2 expressing a c-erbB-2-related antigen, said method
- 3 comprising:
- 4 administering to a patient harboring the tumor a
- 5 tumor inhibiting amount of a therapeutic agent
- 6 comprising a single-chain Fv of claim 1 and at least a
- 7 first moiety peptide bonded thereto, said first moiety
- 8 having the ability to limit the proliferation of a
- 9 tumor cell.
- 1 18. The method of claim 17 wherein said first moiety
- 2 comprises a cell toxin or a toxic fragment thereof.
- 1 19. The method of claim 17 wherein said first moiety
- 2 comprises a radioisotope sufficiently radioactive to
- 3 inhibit proliferation of said tumor cell.
- 1 20. A DNA sequence encoding the polypeptide chain of
- 2 claim 10.



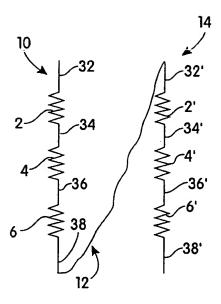
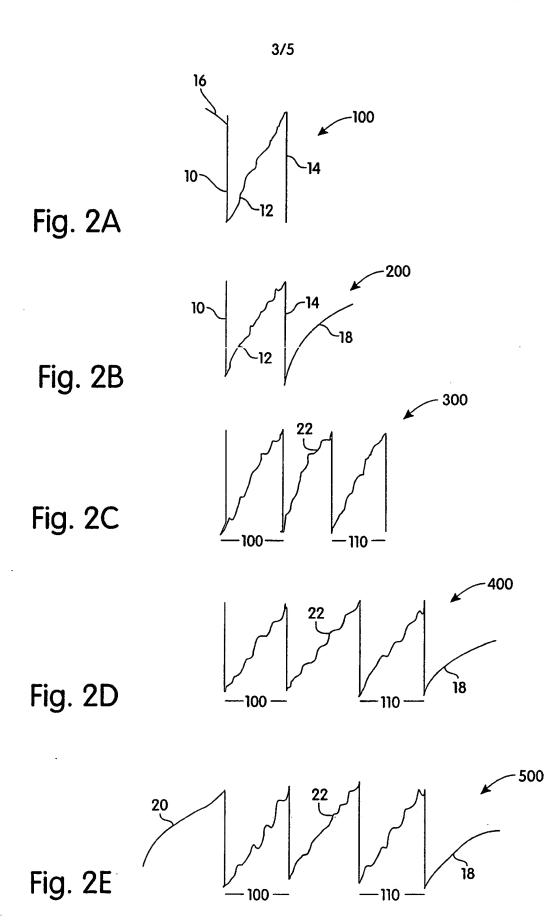


Fig. 1B



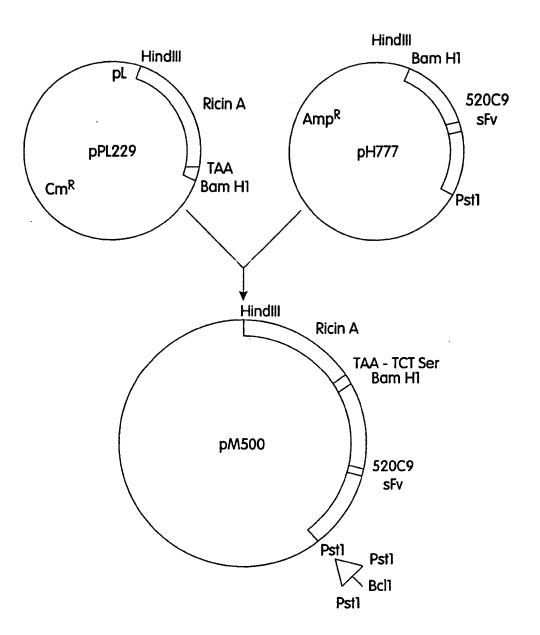
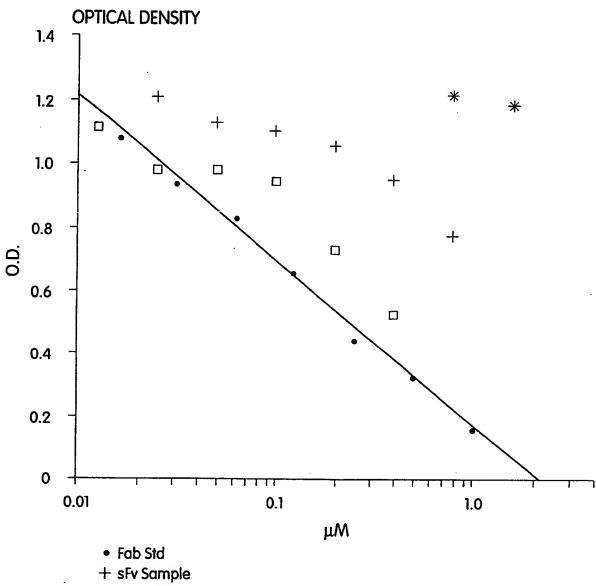


Fig. 3



- □ sFv, Bound and eluted
- * sFv, Unbound and flow through

Fig. 4